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(71) Applicant: **Ajinomoto Co., Inc.**
Tokyo 104 (JP)

(72) Inventors:
- **Hirano, Seiko,**
  Ajinomot Co., Inc.
  Kawasaki-shi, Kanagawa 210 (JP)
- **Sugimoto, Masakazu,**
  Ajinomoto Co., Inc.
  Kawasaki-shi, Kanagawa 210 (JP)
- **Nakano, Eiichi,**
  Ajinomoto Co., Inc.
  Kawasaki-shi, Kanagawa 210 (JP)
- **Izui, Masako,**
  Ajinomoto Co., Inc.
  Kawasaki-shi, Kanagawa 210 (JP)
- **Hayakawa, Atsushi,**
  Ajinomoto Co., Inc.
  Kawasaki-shi, Kanagawa 210 (JP)
- **Yoshihara, Yasuhiko,**
  Ajinomoto Co., Inc.
  Kawasaki-shi, Kanagawa 210 (JP)
- **Nakamatsu, Tsuyoshi,**
  Ajinomoto Co., Inc.
  Kawasaki-shi, Kanagawa 210 (JP)

(74) Representative:
Kolb, Helga, Dr. Dipl.-Chem. et al
Hoffmann Eitle,
Patent- und Rechtsanwälte,
Arabellastrasse 4
81925 München (DE)

(54) Method of producing L-lysine

(57) A coryneform bacterium in which a DNA coding for a diaminopimelate decarboxylase and a DNA coding for a diaminopimelate dehydrogenase are enhanced is cultivated in a medium to allow L-lysine to be produced and accumulated in a culture, and L-lysine is collected from the culture.

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Description

BACKGROUND OF THE INVENTION

The present invention relates to a method for producing L-lysine by cultivating a microorganism obtained by modifying a coryneform bacterium used for fermentative production of amino acids or the like by means of a technique based on genetic engineering.

L-Lysine, which is used as a fodder additive, is usually produced by a fermentative method by using an L-lysine-producing mutant strain belonging to the coryneform bacteria. Various L-lysine-producing bacteria known at present are those created by artificial mutation starting from wild type strains belonging to the coryneform bacteria.

As for the coryneform bacteria, there are disclosed a vector plasmid which is autonomously replicable in bacterial cells and has a drug resistance marker gene (see United States Patent No. 4,514,502), and a method for introducing a gene into bacterial cells (for example, Japanese Patent Application Laid-open No. 2-207791). There is also disclosed a possibility for breeding an L-threonine- or L-isoleucine-producing bacterium by using the techniques as described above (see United States Patent Nos. 4,452,890 and 4,442,208). As for breeding of an L-lysine-producing bacterium, a technique is known, in which a gene participating in L-lysine biosynthesis is incorporated into a vector plasmid to amplify the gene in bacterial cells (for example, Japanese Patent Application Laid-open No. 56-160997).

Known genes for L-lysine biosynthesis include, for example, a dihydrodipicolinate reductase gene (Japanese Patent Application Laid-open No. 7-75578) and a diaminopimelate dehydrogenase gene (Ishino, S. et al., Nucleic Acids Res., 15, 3917 (1987)), which are cloned genes which participate in L-lysine biosynthesis, as well as a phosphoenolpyruvate carboxylase gene (Japanese Patent Application Laid-open No. 60-87788), a dihydrodipicolinate synthase gene (Japanese Patent Publication No. 6-55149), and a diaminopimelate decarboxylase gene (Japanese Patent Application Laid-open No. 60-62994), amplification of which affects L-lysine productivity.

As described above, certain successful results to improve L-lysine productivity have been obtained by means of amplification of genes for the L-lysine biosynthesis. However, amplification of some genes decreases growth speed of bacteria although the amplification improves L-lysine productivity, resulting in decrease of rate of L-lysine production.

No case has been reported in which growth is intended to be improved by enhancing a gene for L-lysine biosynthesis as well. In the present circumstances, no cafe is known for the coryneform bacteria, in which anyone has succeeded in remarkable improvement in L-lysine yield without restraining growth, by combining a plurality of genes for L-lysine biosynthesis.

SUMMARY OF THE INVENTION

An object of the present invention is to improve the L-lysine-producing ability without decreasing the growth speed of a coryneform bacterium, by enhancing a plurality of genes for L-lysine biosynthesis in combination in the coryneform bacteria.

When an objective substance is produced fermentatively by using a microorganism, the production speed, as well as the yield of the objective substance relative to an introduced material, is an extremely important factor. An objective substance may be produced remarkably inexpensively by increasing the production speed per a unit of fermentation equipment. Accordingly, it is industrially extremely important that the fermentative yield and the production speed are compatible with each other. The present invention proposes a solution for the problem as described above in order to fermentatively produce L-lysine by using a coryneform bacterium.

The principle of the present invention is based on the fact that the growth of a coryneform bacterium can be improved, and the L-lysine-producing speed thereof can be improved by enhancing both of a DNA sequence coding for a diaminopimelate dehydrogenase (a diaminopimelate dehydrogenase is hereinafter referred to as "DDC", and a gene coding for a DDC protein is hereinafter referred to as "lysA", if necessary), and a DNA sequence coding for a diaminopimelate decarboxylase (a diaminopimelate decarboxylase is hereinafter referred to as "DDH", and a gene coding for a DDH protein is hereinafter referred to as "ddh", if necessary) compared with the case in which these DNA sequences are each enhanced singly.

Namely, the present invention lies in a recombinant DNA autonomously replicable in cells of coryneform bacteria, comprising a DNA sequence coding for a diaminopimelate dehydrogenase and a DNA sequence coding for a diaminopimelate decarboxylase.

In another aspect, the present invention provides a coryneform bacterium harboring an enhanced DNA sequence coding for a diaminopimelate dehydrogenase and an enhanced DNA sequence coding for a diaminopimelate decarboxylase.

In still another aspect, th present invention provides a method for producing L-lysin comprising the steps of cultivating the coryneform bacterium described above in a medium to allow L-lysine to be produced and accumulated in a culture, and collecting L-lysine from the culture.

The coryneform bacteria referred to in the present invention are a group of microorganisms as defined in Bergey's

Manual of Determinativ Bacteriology, 8th ed., p. 599 (1974), which are aerobic Gram-positive non-acid-fast rods having no spore-forming ability. The coryneform bacteria include bacteria belonging to the genus Corynebacterium, bacteria belonging to the genus Brevibacterium having been hitherto classified into the genus Brevibacterium but united as bacteria belonging to the genus Corynebacterium at present, and bacteria belonging to the genus Brevibacterium closely relative to bact ria belonging to the genus Corynebacterium.

According to the present invention, the L-lysine-producing ability of coryneform bacteria can be improved, and the growth speed can be also improved. The present invention can be applied to ordinary L-lysine-producing bacteria as well as strains with high L-lysine productivity.

BRIEF EXPLANATION OF DRAWINGS

Fig. 1 illustrates a process of construction of a plasmid p299LYSA carrying lysA.
Fig. 2 illustrates a process of construction of a plasmid pLYSAB carrying lysA and Brevi.-ori.
Fig. 3 illustrates a process of construction of a plasmid pPK4D carrying ddh and Brevi.-ori.
Fig. 4 illustrates a process of construction of a plasmid p399DL carrying ddh and lysA.
Fig. 5 illustrates a process of construction of a plasmid pDL carrying ddh, lysA and Brevi.-ori.
Fig. 6 illustrates a process of construction of plasmids p399AKYB and p399AK9B each carrying mutant lysC.
Fig. 7 illustrates a process of construction of a plasmid pDPRB carrying dapB and Brevi.-ori.
Fig. 8 illustrates a process of construction of a plasmid pDPSB carrying dapA and Brevi.-ori.
Fig. 9 illustrates a process of construction of a plasmid pCRCAB carrying lysC, dapB and Brevi.-ori.
Fig. 10 illustrates a process of construction of a plasmid pCB carrying mutant lysC, dapB, and Brevi.-ori.
Fig. 11 illustrates a process of construction of a plasmid pAB carrying dapA, dapB and Brevi.-ori.
Fig. 12 illustrates a process of construction of a plasmid pCAB carrying mutant lysC, dapA, dapB, and Brevi.-ori.
Fig. 13 illustrates a process of construction of a plasmid pCABL carrying mutant lysC, dapA, dapB, lysA, and Brevi.-ori.
Fig. 14 illustrates a process of construction of a plasmid pCABDL carrying mutant lysC, dapA, dapB, ddh, lysA, and Brevi.-ori.

DETAILED DESCRIPTION OF THE INVENTION

The present invention will be explained in detail below.

(1) Preparation of genes for L-lysine biosynthesis used for the present invention

The genes for L-lysine biosynthesis used in the present invention can be obtained respectively by preparing chromosomal DNA from a bacterium as a DNA donor, constructing a chromosomal DNA library by using a plasmid vector or the like, selecting a strain harboring a desired gene from the library, and recovering, from the selected strain, recombinant DNA into which the gene has been inserted. The DNA donor for the gene for L-lysine biosynthesis used in the present invention is not specifically limited provided that the desired gene for L-lysine biosynthesis expresses an enzyme protein which functions in cells of coryneform bacteria. However, the DNA donor is preferably a coryneform bacterium.

Both of the genes of lysA and ddh originating from coryneform bacteria have known sequences. Accordingly, they can be obtained by performing amplification in accordance with the polymerase chain reaction method (PCR; see White, T. J. et al., Trends Genet., 5, 185 (1989)).

Each of the genes for L-lysine biosynthesis used in the present invention is obtainable in accordance with certain methods as exemplified below.

(1) Preparation of mutant lysA

lysA can be isolated from chromosome of a coryneform bacterium by preparing chromosomal DNA in accordance with, for example, a method of Saito and Miura (H. Saito and K. Miura, Biochem. Biophys. Acta, 72, 619 (1963)), and amplifying lysA in accordance with the polymerase chain reaction method (PCR; see White, T. J. et al., Trends Genet., 5, 185 (1989)). The DNA donor is not specifically limited, however, it is exemplified by Brevibacterium lactofermentum ATCC 13869 strain.

In the coryn form bacteria, lysA forms an operon together with argS (arginyl-tRNA synthase gene), and lysA exists downstream from argS. Expression of lysA is regulated by a promoter existing upstream from argS (see Journal of Bacteriology, Nov., 7356-7362 (1993)). DNA sequences of these genes are known for Corynebacterium glutamicum (see Molecular Microbiology, 4(11), 1819-1830 (1990); Molecular and General Genetics, 212, 112-119 (1988)), on the basis of which DNA primers for PCR can be prepared. Such DNA primers are specifically exemplified by DNA's of 23-mers

respectively having nucleotide sequences shown in SEQ ID NO: 1 in Sequence Listing (corresponding to nucleotide numbers 11 to 33 in a nucleotide sequence described in Molecular Microbiology, 4(11), 1819-1830 (1990)) and SEQ ID NO: 2 (corresponding to nucleotide numbers 1370 to 1392 in a nucleotide sequence described in Molecular and General Genetics, 212, 112-119 (1988)).

In Example described later on, a DNA fragment containing a promoter, argS, and lysA was used in order to enhance lysA. However, argS is not essential for the present invention. It is allowable to use a DNA fragment in which lysA is ligated just downstream from a promoter.

A nucleotide sequence of a DNA fragment containing argS and lysA, and an amino acid sequence deduced to be encoded by the nucleotide sequence are exemplified in SEQ ID NO: 3. An example of an amino acid sequence encoded by argS is shown in SEQ ID NO: 4, and an example of an amino acid sequence encoded by lysA is shown in SEQ ID NO: 5. In addition to DNA fragments coding for these amino acid sequences, the present invention can equivalently use DNA fragments coding for amino acid sequences substantially the same as the amino acid sequence shown in SEQ ID NO: 5, namely amino acid sequences having mutation based on, for example, substitution, deletion, or insertion of one or more amino acids provided that there is no substantial influence on the DDC activity.

DNA can be synthesized in accordance with an ordinary method by using DNA synthesizer model 380B produced by Applied Biosystems and using the phosphoamidite method (see Tetrahedron Letters (1981), 22, 1859). PCR can be performed by using DNA Thermal Cycler Model PJ2000 produced by Takara Shuzo, and using Taq DNA polymerase in accordance with a method designated by the supplier.

It is preferred that lysA amplified by PCR is ligated with vector DNA autonomously replicable in cells of E. coli and/or coryneform bacteria to prepare recombinant DNA, and the recombinant DNA is introduced into cells of E. coli beforehand. Such provision makes following operations easy. The vector autonomously replicable in cells of E. coli is preferably a plasmid vector which is preferably autonomously replicable in cells of a host, including, for example, pUC19, pUC18, pBR322, pHSG299, pHSG399, pHSG398, and RSF1010. When a DNA fragment having an ability to allow a plasmid to be autonomously replicable in coryneform bacteria is inserted into these vectors, they can be used as a so-called shuttle vector autonomously replicable in both E. coli and coryneform bacteria.

Such a shuttle vector includes the followings. Microorganisms harboring each of vectors and accession numbers in international depositary authorities are shown in parentheses.

pHC4: Escherichia coli AJ12617 (FERM BP-3532)
pAJ655: Escherichia coli AJ11882 (FERM BP-136) Corynebacterium glutamicum SR8201 (ATCC 39135)
pAJ1844: Escherichia coli AJ11883 (FERM BP-137) Corynebacterium glutamicum SR8202 (ATCC 39136)
pAJ611: Escherichia coli AJ11884 (FERM BP-138)
pAJ3148: Corynebacterium glutamicum SR8203 (ATCC 39137)
pAJ440: Bacillus subtilis AJ11901 (FERM BP-140)

These vectors are obtainable from the deposited microorganisms as follows. Cells collected at a logarithmic growth phase were lysed by using lysozyme and SDS, followed by separation from a lysate by centrifugation at 30,000 × g to obtain a supernatant. Polyethylene glycol is added to the supernatant, followed by fractionation and purification by means of cesium chloride-ethidium bromide equilibrium density gradient centrifugation.

E. coli can be transformed by introducing a plasmid in accordance with, for example, a method of D. M. Morrison (Methods in Enzymology, 68, 326 (1979)) or a method in which recipient cells are treated with calcium chloride to increase permeability for DNA (Mandel, M. and Higa, A., J. Mol. Biol., 53, 159 (1970)).

(2) Preparation of ddh

A DNA fragment containing ddh can be prepared from chromosome of a coryneform bacterium by means of PCR. Th DNA donor is not specifically limited, however, it is exemplified by Brevibacterium lactofermentum ATCC 13869 strain.

A DDH gene is known for Corynebacterium glutamicum (Ishino, S. et al., Nucleic Acids Res., 15, 3917 (1987)), on the basis of which primers for PCR can be prepared. Such DNA primers are specifically exemplified by DNA's of 20-mers respectively having nucleotide sequences shown in SEQ ID NOs: 6 and 7 in Sequence Listing. Synthesis of DNA, PCR, and preparation of a plasmid carrying obtained ddh can be performed in the same manner as those for lysA described above.

A nucleotide sequence of a DNA fragment containing ddh and an amino acid sequence deduced from the nucleotide sequence are illustrated in SEQ ID NO: 8. Only th amino acid sequence is shown in SEQ ID NO: 9. In addition to DNA fragments coding for this amin acid sequence, the pres nt invention can equivalently use DNA fragments coding for amino acid sequences substantially the same as the amino acid sequence shown in SEQ ID NO: 9, namely amino acid sequences having mutation based on, for example, substitution, deletion, or insertion of one or more amino acids provided that there is no substantial influence on the DDH activity.

(2) Recombinant DNA and coryneform bacterium of the present invention

The coryneform bacterium of the present invention harbors a DNA sequence coding for diaminopimelate decarboxylase (lysA) and a DNA sequence coding for diaminopimelate dehydrogenase (ddh) which are enhanced. The term "a DNA sequence is enhanced" herein refers to the fact that the intracellular activity of an enzyme encoded by the DNA sequence is raised by, for example, increasing the copy number of a gene, using a strong promoter, using a gene coding for an enzyme having a high specific activity, or combining these means.

The coryneform bacterium to which the DNA sequences described above is an L-lysine-producing coryneform bacterium, examples of which include L-lysine-producing wild type strains and artificial mutant strains and coryneform bacteria enhanced in L-lysine productivity by genetic engineering. Even if the bacterium has low L-lysine productivity, the L-lysine productivity can be improved by enhancing lysA and ddh. If the bacterium has high L-lysine productivity, the L-lysine production efficiency can be more raised by enhancing lysA and ddh.

(1) L-Lysine-producing strain belonging to coryneform bacteria

Examples of the coryneform bacterium used to introduce lysA and ddh include, for example, the following lysine-producing strains:

Corynebacterium acetoacidonhilum ATCC 13870;
Corynebacterium acetoglutamicum ATCC 15806;
Corynebacterium callunae ATCC 15991;
Corynebacterium glutamicum ATCC 13032;
(Brevibacterium divaricatum) ATCC 14020;
(Brevibacterium lactofermentum) ATCC 13869;
(Corynebacterium lilium) ATCC 15990;
(Brevibacterium flavum) ATCC 14067;
Corynebacterium melassecola ATCC 17965;
Brevibacterium saccharolyticum ATCC 14066;
Brevibacterium immariophilum ATCC 14068;
Brevibacterium roseum ATCC 13825;
Brevibacterium thiogenitalis ATCC 19240;
Microbacterium ammoniaphilum ATCC 15354;
Corynebacterium thermoaminogenes AJ12340 (FERM BP-1539).

Other than the bacterial strains described above, those usable as a host in which lysA and ddh are to be enhanced include, for example, mutant strains having an L-lysine-producing ability derived from the aforementioned strains. Such artificial mutant strains include the followings: S-(2-aminoethyl)-cysteine (hereinafter abbreviated as "AEC") resistant mutant strains (Brevibacterium lactofermentum AJ11082 (NRRL B-11470), Japanese Patent Publication Nos. 56-1914, 56-1915, 57-14157, 57-14158, 57-30474, 58-10075, 59-4993, 61-35840, 62-24074, 62-36673, 5-11958, 7-112437, and 7-112438); mutant strains which require an amino acid such as L-homoserine for their growth (Japanese Patent Publication Nos. 48-28078 and 56-6499); mutant strains which exhibit resistance to AEC and require amino acids such as L-leucine, L-homoserine, L-proline, L-serine, L-arginine, L-alanine, and L-valine (United States Patent Nos. 3,708,395 and 3,825,472); L-lysine-producing mutant strains which exhibit resistance to DL-α-amino-ε-caprolactam, α-amino-lauryllactam, aspartate-analog, sulfa drug, quinoid, and N-lauroylleucine; L-lysine-producing mutant strains which exhibit resistance to inhibitors of oxyaloacetate decarboxylase or respiratory system enzymes (Japanese Patent Application Laid-open Nos. 50-53588, 50-31093, 52-102498, 53-9394, 53-86089, 55-9783, 55-9759, 56-32995 and 56-39778, and Japanese Patent Publication Nos. 53-43591 and 53-1833); L-lysine-producing mutant strains which require inositol or acetic acid (Japanese Patent Application Laid-open Nos. 55-9784 and 56-8692); L-lysine-producing mutant strains which exhibit sensitivity to fluoropyruvic acid or temperature not less than 34°C (Japanese Patent Application Laid-open Nos. 55-9783 and 53-86090); and production mutant strains belonging to the genus Brevibacterium or Corynebacterium which exhibit resistance to ethylene glycol and produce L-lysine (United States Patent No. 4,411,997).

(2) L-Lysine-producing coryneform bacteria having L-lysine productivity enhanced by genetic recombination

Th L-lysine producing speed can be further improved by enhancing lysA and ddh, if the coryn form bacterium has been enhanced in L-lysine production by genetic engineering, for example, by introducing a gene coding for an enzyme having a mutation which causes desensitization in feedback inhibition, wild type of which enzyme is subjected to feedback inhibition among enzymes participating in L-lysine biosynthesis, or by enhancing a gene for L-lysine biosynthesis

other than lysA and ddh.

The coryneform bacterium enhanced in L-lysine productivity includes a coryneform bacterium harboring a DNA sequence coding for an aspartokinase which is desensitized in feedback inhibition by L-lysine and L-threonine (an aspartokinase is hereinafter referred to as "AK", a gene coding for an AK protein is hereinafter referred to as "lysC", and a gene coding for an AK protein which is desensitized in feedback inhibition by L-lysine and L-threonine, if necessary), and an enhanced DNA sequence coding for a dihydrodipicolinate reductase (a dihydrodipicolinate reductase is hereinafter referred to as "DDPR", and a gene coding for a DDPR protein is hereinafter referred to as "dapB", if necessary), and the coryneform bacterium further harboring an enhanced DNA sequence coding for a dihydrodipicolinate synthase (a dihydrodipicolinate synthase is hereinafter referred to as "DDPS", and a gene coding for a DDPS protein is hereinafter referred to as "dapA", if necessary). Each of the genes for L-lysine biosynthesis used in the present invention is obtainable in accordance with certain methods as exemplified below.

(i) Preparation of mutant lysC

A DNA fragment containing mutant lysC can be prepared from a mutant strain in which synergistic feedback inhibition on the AK activity by L-lysine and L-threonine is substantially desensitized (International Publication No. WO 94/25605). Such a mutant strain can be obtained, for example, from a group of cells originating from a wild type strain of a coryneform bacterium subjected to a mutation treatment by applying an ordinary mutation treatment such as ultraviolet irradiation and treatment with a mutating agent such as N-methyl-N'-nitro-N-nitrosoguanidine. The AK activity can be measured by using a method described by Miyajima, R. et al., The Journal of Biochemistry (1968), 63(2), 139-148. The most preferred as such a mutant strain is represented by an L-lysine-producing bacterium AJ3445 (FERM P-1944) derived by a mutation treatment from a wild type strain of Brevibacterium lactofermentum ATCC 13869 (having its changed present name of Corynebacterium glutamicum).

Alternatively, mutant lysC is also obtainable by an in vitro mutation treatment of plasmid DNA containing wild type lysC. In another aspect, information is specifically known on mutation to desensitize synergistic feedback inhibition on AK by L-lysine and L-threonine (International Publication No. WO 94/25605). Accordingly, mutant lysC can be also prepared from wild type lysC on the basis of the information in accordance with, for example, the site-directed mutagenesis method.

A DNA fragment containing lysC can be prepared from chromosome of a coryneform bacterium by means of PCR. DNA primers are exemplified by single strand DNA's of 23-mer and 21-mer having nucleotide sequences shown in SEQ ID NOs: 10 and 11 in Sequence Listing in order to amplify, for example, a region of about 1,643 bp coding for lysC based on a sequence known for Corynebacterium glutamicum (see Molecular Microbiology (1991), 5(5), 1197-1204; Mol. Gen. Genet. (1990), 224, 317-324). Synthesis of DNA, PCR, and preparation of a plasmid carrying obtained lysC can be performed in the same manner as those for lysA described above.

Wild type lysC is obtained when lysC is isolated from an AK wild type strain, while mutant lysC is obtained when lysC is isolated from an AK mutant strain in accordance with the method as described above.

An example of a nucleotide sequence of a DNA fragment containing wild type lysC is shown in SEQ ID NO: 12 in Sequence Listing. An amino acid sequence of α-subunit of a wild type AK protein is deduced from the nucleotide sequence, and is shown in SEQ ID NO: 13 in Sequence Listing together with the DNA sequence. Only the amino acid sequence is shown in SEQ ID NO: 14. An amino acid sequence of β-subunit of the wild type AK protein is deduced from the nucleotide sequence of DNA, and is shown in SEQ ID NO: 15 in Sequence Listing together with the DNA sequence. Only the amino acid sequence is shown in SEQ ID NO: 16. In each of the subunits, GTG is used as an initiation codon, and a corresponding amino acid is represented by methionine. However, this representation refers to methionine, valine, or formylmethionine.

The mutant lysC used in the present invention is not specifically limited provided that it codes for AK in which synergistic feedback inhibition by L-lysine and L-threonine is desensitized. However, the mutant lysC is exemplified by one including mutation in which a 279th alanine residue as counted from the N-terminal is changed into an amino acid residue other than alanine and other than acidic amino acid in the α-subunit, and a 30th alanine residue is changed into an amino acid residue other than alanine and other than acidic amino acid in the β-subunit in the amino acid sequence of the wild type AK. The amino acid sequence of the wild type AK specifically includes the amino acid sequence shown in SEQ ID NO: 14 in Sequence Listing as the α-subunit, and the amino acid sequence shown in SEQ ID NO: 16 in Sequence Listing as the β-subunit.

Those preferred as the amino acid residue other than alanine and other than acidic amino acid include threonine, arginine, cysteine, phenylalanine, proline, serine, tyrosine, and valine residues.

The codon corresponding to an amino acid residue to be substituted is not specifically limited for its type provided that it codes for the amino acid residue. It is assumed that th amino acid sequence of possessed wild typ AK may slightly differ depending on the difference in bacterial species and bacterial strains. AK's, which have mutation based on, for example, substitution, deletion, or insertion of one or more amino acid residues at one or more positions irrelevant to the enzyme activity as described above, can be also used for the present invention. Other AK's, which have

mutation based on, for example, substitution, deletion, or insertion of other one or more amino acid residues, can be also used provided that no influence is substantially exerted on the AK activity, and on the desensitization of synergistic feedback inhibition by L-lysine and L-threonine.

An AJ12691 strain obtained by introducing a mutant lysC plasmid p399AK9B into an AJ12036 strain (FERM BP-734) as a wild type strain of Brevibacterium lactofermentum has been deposited on April 10, 1992 under an accession number of FERM P-12918 in National Institute of Bioscience and Human Technology of Agency of Industrial Science and Technology of Ministry of International Trade and Industry (postal code: 305, 1-3, Higashi 1-chome, Tsukuba-shi, Ibaraki-ken, Japan), transferred to international deposition based on the Budapest Treaty on February 10, 1995, and deposited under an accession number of FERM BP-4999.

(ii) Preparation of dapB

A DNA fragment containing dapB can be prepared from chromosome of a coryneform bacterium by means of PCR. The DNA donor is not specifically limited, however, it is exemplified by Brevibacterium lactofermentum ATCC 13869 strain.

A DNA sequence coding for DDPR is known for Brevibacterium lactofermentum (Journal of Bacteriology, 175(9), 2743-2749 (1993)), on the basis of which DNA primers for PCR can be prepared. Such DNA primers are specifically exemplified by DNA's of 23-mers respectively having nucleotide sequences shown in SEQ ID NOs: 17 and 18 in Sequence Listing. Synthesis of DNA, PCR, and preparation of a plasmid carrying obtained dapB can be performed in the same manner as those for lysC described above.

A nucleotide sequence of a DNA fragment containing dapB and an amino acid sequence deduced from the nucleotide sequence are illustrated in SEQ ID NO: 19. Only the amino acid sequence is shown in SEQ ID NO: 20. In addition to DNA fragments coding for this amino acid sequence, the present invention can equivalently use DNA fragments coding for amino acid sequences substantially the same as the amino acid sequence shown in SEQ ID NO: 20, namely amino acid sequences having mutation based on, for example, substitution, deletion, or insertion of one or more amino acids provided that there is no substantial influence on the DDPR activity.

A transformant strain AJ13107 obtained by introducing a plasmid pCRDAPB carrying dapB obtained in Example described later on into E. coli JM109 strain has been internationally deposited since May 26, 1995 under an accession number of FERM BP-5114 in National Institute of Bioscience and Human Technology of Agency of Industrial Science and Technology of Ministry of International Trade and Industry (postal code: 305, 1-3, Higashi 1-chome, Tsukuba-shi, Ibaraki-ken, Japan) based on the Budapest Treaty.

(iii) Preparation of dapA

A DNA fragment containing dapA can be prepared from chromosome of a coryneform bacterium by means of PCR. The DNA donor is not specifically limited, however, it is exemplified by Brevibacterium lactofermentum ATCC 13869 strain.

A DNA sequence coding for DDPS is known for Corynebacterium glutamicum (see Nucleic Acids Research, 18(21), 6421 (1990); EMBL accession No. X53993), on the basis of which DNA primers for PCR can be prepared. Such DNA primers are specifically exemplified by DNA's of 23-mers respectively having nucleotide sequences shown in SEQ ID NOs: 21 and 22 in Sequence Listing. Synthesis of DNA, PCR, and preparation of a plasmid carrying obtained dapA can be performed in the same manner as those for lysC described above.

A nucleotide sequence of a DNA fragment containing dapA and an amino acid sequence deduced from the nucleotide sequence are exemplified in SEQ ID NO: 23. Only the amino acid sequence is shown in SEQ ID NO: 24. In addition to DNA fragments coding for this amino acid sequence, the present invention can equivalently use DNA fragments coding for amino acid sequences substantially the same as the amino acid sequence shown in SEQ ID NO: 24, namely amino acid sequences having mutation based on, for example, substitution, deletion, or insertion of one or more amino acids provided that there is no substantial influence on the DDPS activity.

A transformant strain AJ13106 obtained by introducing a plasmid pCRDAPA carrying dapA obtained in Example described later on into E. coli JM109 strain has been internationally deposited since May 26, 1995 under an accession number of FERM BP-5113 in National Institute of Bioscience and Human Technology of Agency of Industrial Science and Technology of Ministry of International Trade and Industry (postal code: 305, 1-3, Higashi 1-chome, Tsukuba-shi, Ibaraki-ken, Japan) based on the Budapest Treaty.

In a specified embodiment, in order to enhance lysA and ddh in the L-lysine-producing coryneform bacterium as described above, the genes are introduced into the host by using a plasmid vector, transposon or phage vector or the like. Upon the introduction, it is expected to make enhancement to some extent even by using a low copy type vector. However, it is preferred to use a multiple copy type vector. Such a vector includes, for example, plasmid vectors, pAJ655, pAJ1844, pAJ611, pAJ3148, and pAJ440 described above. Besides, transposons derived from coryneform bacteria are described in International Publication Nos. WO 92/02627 and WO 93/18151; European Patent Publication

No. 445385; Japanese Patent Application Laid-open No. 6-46867; Vertes, A. A. et al., Mol. Microbiol., 11, 739-746 (1994); Bonamy, C., et al., Mol. Microbiol., 14, 571-581 (1994); Vertes, A. A. et al., Mol. Gen. Genet., 245, 397-405 (1994); Jagar, W. et al., FEMS Microbiology Letters, 126, 1-6 (1995); Japanese Patent Application Laid-open Nos. 7-107976 and 7-327680 and the like.

A coryneform bacterium enhanced in lysA and ddh according to the present invention can be obtained, for example, by introducing, into a host coryneform bacterium, a recombinant DNA containing lysA and ddh and being autonomously replicable in cells of coryneform bacteria. The recombinant DNA can be obtained, for example, by inserting lysA and ddh into a vector such as plasmid vector, transposon or phage vector as described above.

Each of the genes of lysA and ddh may be successively introduced into the host by using different vectors respectively. Alternatively, two species of the genes may be introduced together by using a single vector. When different vectors are used, the genes may be introduced in any order, however, it is preferred to use vectors which have a stable sharing and harboring mechanism in the host, and which are capable of coexisting with each other.

In the case in which a plasmid is used as a vector, the recombinant DNA can be introduced into the host in accordance with an electric pulse method (Sugimoto et al., Japanese Patent Application Laid-open No. 2-207791). Amplification of a gene using transposon can be performed by introducing a plasmid carrying a transposon into the host cell and inducing transposition of the transposon.

Also, when mutant lysC, dapA and dapB are introduced into coryneform bacterium, each of the genes and lysA and ddh may be successively introduced into the host by using different vectors respectively or, alternatively, two or more species of the genes may be introduced together by using a single vector.

(3) Method for producing L-lysine

L-Lysine can be efficiently produced by cultivating, in an appropriate medium, the coryneform bacterium comprising the enhanced genes for L-lysine biosynthesis as described above to allow L-lysine to be produced and accumulated in a culture, and collecting L-lysine from the culture.

The medium to be used is exemplified by an ordinary medium containing a carbon source, a nitrogen source, inorganic ions, and optionally other organic components.

As the carbon source, it is possible to use sugars such as glucose, fructose, sucrose, molasses, and starch hydrolysate; and organic acids such as fumaric acid, citric acid, and succinic acid.

As the nitrogen source, it is possible to use inorganic ammonium salts such as ammonium sulfate, ammonium chloride, and ammonium phosphate; organic nitrogen such as soybean hydrolysate; ammonia gas; and aqueous ammonia.

As organic trace nutrient sources, it is desirable to contain required substances such as vitamin B_1 and L-homoserine or yeast extract or the like in appropriate amounts. Other than the above, potassium phosphate, magnesium sulfate, iron ion, manganese ion and so on are added in small amounts, if necessary.

Cultivation is preferably carried out under an aerobic condition for about 30 to 90 hours. The cultivation temperature is pr ferably controlled at 25°C to 37°C, and pH is preferably controlled at 5 to 8 during cultivation. Inorganic or organic, acidic or alkaline substances, or ammonia gas or the like can be used for pH adjustment. L-lysine can be collected from a culture by combining an ordinary ion exchange resin method, a precipitation method, and other known methods.

EXAMPLES

The present invention will be more specifically explained below with reference to Examples.

Example 1: Preparation of lysA from Brevibacterium lactofermentum

(1) Preparation of lysA and construction of plasmid carrying lysA

A wild type strain ATCC 13869 of Brevibacterium lactofermentum was used as a chromosomal DNA donor. Chromosomal DNA was prepared from the ATCC 13869 strain in accordance with an ordinary method. A DNA fragment containing argS, lysA, and a promoter of an operon containing them was amplified from the chromosomal DNA in accordance with PCR. As for DNA primers used for amplification, synthetic DNA's of 23-mers having nucleotide sequences shown in SEQ ID NOs: 1 and 2 in Sequence Listing respectively were used in order to amplify a region of about 3.6 kb coding for arginyl-tRNA synthase and DDC on the basis of a sequence known for Corynebacterium glutamicum (see Molecular Microbiology, 4(11), 1819-1830 (1990); Molecular and General Genetics, 212, 112-119 (1988)).

DNA was synthesized in accordance with an ordinary method by using DNA synthesizer model 380B produced by Applied Biosystems and using the phosphoamidite method (see Tetrahedron Letters (1981), 22, 1859).

The gene was amplified by PCR by using DNA Thermal Cycler Model PJ2000 produced by Takara Shuzo, and using Taq DNA polymerase in accordance with a method designated by the supplier. pHSG399 was used as a cloning

vector for the amplified gene fragment of 3,579 bp. pHSG399 was digested with a restriction enzyme SmaI (produced by Takara Shuzo), and was ligated with the DNA fragment containing amplified lysA. A plasmid obtained as described above, which had lysA originating from ATCC 13869, was designated as p399LYSA.

A DNA fragment containing lysA was extracted by digesting p399LYSA with KpnI (produced by Takara Shuzo) and BamHI (produced by Takara Shuzo). This DNA fragment was ligated with pHSG299 having been digested with KpnI and BamHI. An obtained plasmid was designated as p299LYSA. The process of construction of p299LYSA is shown in Fig. 1.

A DNA fragment (hereinafter referred to as "Brevi.-ori") having an ability to make a plasmid autonomously replicable in bacteria belonging to the genus Corynebacterium was introduced into p299LYSA to prepare plasmids carrying lysA autonomously replicable in bacteria belonging to the genus Corynebacterium. Brevi.-ori was prepared from a plasmid vector pHK4 containing Brevi.-ori and autonomously replicable in cells of both Escherichia coli and bacteria belonging to the genus Corynebacterium. pHK4 was constructed by digesting pHC4 with KpnI (produced by Takara Shuzo) and BamHI (produced by Takara Shuzo), extracting a Brevi.-ori fragment, and ligating it with pHSG298 having been also digested with KpnI and BamHI (see Japanese Patent Application Laid-open No. 5-7491). pHK4 gives kanamycin resistance to a host. Escherichia coli HB101 harboring pHK4 was designated as Escherichia coli AJ13136, and deposited on August 1, 1995 under an accession number of FERM BP-5186 in National Institute of Bioscience and Human Technology of Agency of Industrial Science and Technology of Ministry of International Trade and Industry (postal code: 305, 1-3, Higashi 1-chome, Tsukuba-shi, Ibaraki-ken, Japan).

pHK4 was digested with a restriction enzyme BamHI, and cleaved ends were blunted. Blunt end formation was performed by using DNA Blunting kit (produced by Takara Shuzo) in accordance with a designated method. After the blunt end formation, a phosphorylated KpnI linker (produced by Takara Shuzo) was ligated to make modification so that the DNA fragment corresponding to the Brevi.-ori portion might be excised from pHK4 by digestion with only KpnI. This plasmid was digested with KpnI, and the generated Brevi.-ori DNA fragment was ligated with p299LYSA having been also digested with KpnI to prepare a plasmid carrying lysA autonomously replicable in coryneform bacteria. The prepared plasmid was designated as pLYSAB. The process of construction of pLYSAB is shown in Fig. 2.

(2) Determination of nucleotide sequence of lysA from Brevibacterium lactofermentum

Plasmid DNA of p299LYSA was prepared, and nucleotide sequence determination was performed in accordance with a method of Sanger et al. (for example, F. Sanger et al., Proc. Natl. Acad. Sci., 74, 5463 (1977)). A determined nucleotide sequence and an amino acid sequence deduced to be encoded by the nucleotide sequence are shown in SEQ ID NO: 3. Concerning the nucleotide sequence, an amino acid sequence encoded by argS and an amino acid sequence encoded by lysA are shown in SEQ ID NOs: 4 and 5 respectively.

Example 2: Preparation of ddh from Brevibacterium lactofermentum

A ddh gene was obtained by amplifying the ddh gene from chromosomal DNA of Brevibacterium lactofermentum ATCC 13869 in accordance with the PCR method by using two oligonucleotide primers (SEQ ID NOs: 6, 7) prepared on the basis of a known nucleotide sequence of a ddh gene of Corynebacterium glutamicum (Ishino, S. et al., Nucleic Acids Res., 15, 3917 (1987)). An obtained amplified DNA fragment was digested with EcoT22I and AvaI, and cleaved ends were blunted. After that, the fragment was inserted into a SmaI site of pMW119 to obtain a plasmid pDDH.

Next, pDDH was digested with SalI and EcoRI, followed by blunt end formation. After that, an obtained fragment was ligated with pUC18 having been digested with SmaI. A plasmid thus obtained was designated as pUC18DDH.

Brevi.-ori was introduced into pUC18DDH to construct a plasmid carrying ddh autonomously replicable in coryneform bacteria. pHK4 was digested with restriction enzymes KpnI and BamHI, and cleaved ends were blunted. Blunt end formation was performed by using DNA Blunting kit (produced by Takara Shuzo) in accordance with a designated method. After the blunt end formation, a phosphorylated PstI linker (produced by Takara Shuzo) was ligated so that it was inserted into a PstI site of pHSG299. A plasmid constructed as described above was designated as pPK4. Next, pUC18DDH was digested with XbaI and KpnI, and a generated fragment was ligated with pPK4 having been digested with KpnI and XbaI. Thus a plasmid carrying ddh autonomously replicable in coryneform bacteria was constructed. This plasmid was designated as pPK4D. The process of construction of pPK4D is shown in Fig. 3.

Example 3: Construction of Plasmid Carrying Both of ddh and lysA

The plasmid pUC18DDH carrying ddh was digested with EcoRI and then blunt-ended and further digested with XbaI to extract a DNA fragment containing ddh. This ddh fragment was ligated with the plasmid p399LYSA carrying lysA having been digested with BamHI and then blunt-ended and further having been digested with XbaI. An obtained plasmid was designated as p399DL. The process of construction of p399DL is shown in Fig. 4.

Next, Brevi.-ori was introduced into p399DL. pHK4 was digested with XbaI and BamHI, and cleaved ends were

blunted. After the blunt end formation, a phosphorylated XbaI linker was ligated to make modification so that the DNA fragment corresponding to the Brevi.-ori portion might be excised from pHK4 by digestion with only XbaI. This plasmid was digested with XbaI, and the generated Brevi.-ori DNA fragment was ligated with p399DL having been also digested with XbaI to construct a plasmid containing ddh and lysA autonomously replicable in coryneform bacteria. The constructed-plasmid was designated as pDL. The process of construction of pDL is shown in Fig. 5.

Example 4: Preparation of Mutant lysC, dapA and dapB from Brevibacterium lactofermentum

(1) Preparation of Wild Type lysC Gene and Mutant lysC Gene from Brevibacterium lactofermentum

(1) Preparation of wild type and mutant lysC's and preparation of plasmids carrying them

A strain of Brevibacterium lactofermentum ATCC 13869, and an L-lysine-producing mutant strain AJ3445 (FERM P-1944) obtained from the ATCC 13869 strain by a mutation treatment were used as chromosomal DNA donors. The AJ3445 strain had been subjected to mutation so that lysC was changed to involve substantial desensitization from concerted inhibition by lysine and threonine (Journal of Biochemistry, 68, 701-710 (1970)).

A DNA fragment containing lysC was amplified from chromosomal DNA in accordance with the PCR method (polymerase chain reaction; see White, T. J. et al., Trends Genet., 5, 185 (1989)). As for DNA primers used for amplification, single strand DNA's of 23-mer and 21-mer having nucleotide sequences shown in SEQ ID NOs: 10 and 11 were synthesized in order to amplify a region of about 1,643 bp coding for lysC on the basis of a sequence known for Corynebacterium glutamicum (see Molecular Microbiology (1991), 5(5), 1197-1204; and Mol. Gen. Genet. (1990), 224, 317-324).

An amplified gene fragment of 1,643 kb was confirmed by agarose gel electrophoresis. After that, the fragment excised from the gel was purified in accordance with an ordinary method, and it was digested with restriction enzymes NruI (produced by Takara Shuzo) and EcoRI (produced by Takara Shuzo).

pHSG399 (see Takeshita, S. et al., Gene (1987), 61, 63-74) was used as a cloning vector for the gene fragment. pKSG399 was digested with restriction enzymes SmaI (produced by Takara Shuzo) and EcoRI, and it was ligated with the amplified lysC fragment. DNA was ligated by using DNA ligation kit (produced by Takara Shuzo) in accordance with a designated method. Thus plasmids were prepared, in which the lysC fragments amplified from chromosomes of Brevibacterium lactofermentum were ligated with pHSG399 respectively. A plasmid carrying lysC from ATCC 13869 (wild type strain) was designated as p399AKY, and a plasmid carrying lysC from AJ3463 (L-lysine-producing bacterium) was designated as p399AK9.

Brevi.-ori was introduced into the prepared p399AKY and p399AK9 respectively to construct plasmids carrying lysC autonomously replicable in coryneform bacteria.

pHK4 was digested with restriction enzymes KpnI and BamHI (produced by Takara Shuzo), and cleaved ends were blunted. Blunt end formation was performed by using DNA Blunting kit (produced by Takara Shuzo) in accordance with a designated method. After the blunt end formation, a phosphorylated BamHI linker (produced by Takara Shuzo) was ligated to make modification so that the DNA fragment corresponding to the Brevi.-ori portion might be excised from pHK4 by digestion with only BamHI. This plasmid was digested with BamHI, and the generated Brevi.-ori DNA fragment was ligated with p399AKY and p399AK9 having been also digested with BamHI to prepare plasmids carrying lysC autonomously replicable in coryneform bacteria.

A plasmid carrying the wild type lysC gene originating from p399AKY was designated as p399AKYB, and a plasmid carrying the mutant lysC gene originating from p399AK9 was designated as p399AK9B. The process of construction of p399AK9B and p399AKYB is shown in Fig. 6. A strain AJ12691 obtained by introducing the mutant lysC plasmid p399AK9B into a wild type strain of Brevibacterium lactofermentum (AJ12036 strain, FERM BP-734) was deposited on April 10, 1992 under an accession number of FERM P-12918 in National Institute of Bioscience and Human Technology of Agency of Industrial Science and Technology of Ministry of International Trade and Industry (postal code: 305, 1-3, Higashi 1-chome, Tsukuba-shi, Ibaraki-ken, Japan), transferred to international deposition based on the Budapest Treaty on February 10, 1995, and deposited under an accession number of FERM BP-4999.

(2) Determination of nucleotide sequences of wild type lysC and mutant lysC from Brevibacterium lactofermentum

The plasmid p399AKY containing the wild type lysC and the plasmid p399AK9 containing the mutant lysC were prepared from the respective transformants to determine nucleotide sequences of the wild type and mutant lysC's. Nucleotide sequence determination was performed in accordance with a method of Sanger et al. (for example, F. Sanger et al., Proc. Natl. Acad. Sci., 74, 5463 (1977)).

The nucleotide sequence of wild type lysC encoded by p399AKY is shown in SEQ ID NO: 12 in Sequence Listing. On the other hand, the nucleotide sequence of mutant lysC encoded by p399AK9 had only mutation of one nucleotide such that 1051st G was changed into A in SEQ ID NO: 12 as compared with wild type lysC. It is known that lysC of

Corynebacterium glutamicum has two subunits (α, β) encoded in an identical reading frame on an identical DNA strand (see Kalinowski, J. et al., Molecular Microbiology (1991) 5(5), 1197-1204). Judging from homology, it is expected that the gene sequenced herein also has two subunits (α, β) encoded in an identical reading frame on an identical DNA strand.

An amino acid sequence of the α-subunit of the wild type AK protein deduced from the nucleotide sequence of DNA is shown in SEQ ID NO: 13 together with the DNA sequence. Only the amino acid sequence is shown in SEQ ID NO: 14. An amino acid sequence of the β-subunit of the wild type AK protein deduced from the nucleotide sequence of DNA is shown in SEQ ID NO: 15 together with DNA. Only the amino acid sequence is shown in SEQ ID NO: 16. In each of the subunits, GTG is used as an initiation codon, and a corresponding amino acid is represented by methionine. However, this representation refers to methionine, valine, or formylmethionine.

On the other hand, mutation on the sequence of mutant lysC means occurrence of amino acid residue substitution such that a 279th alanine residue of the α-subunit is changed into a threonine residue, and a 30th alanine residue of the β-subunit is changed into a threonine residue in the amino acid sequence of the wild type AK protein (SEQ ID NOs: 14, 16).

(2) Preparation of dapB from Brevibacterium lactofermentum

(1) Preparation of dapB and construction of plasmid carrying dapB

A wild type strain ATCC 13869 of Brevibacterium lactofermentum was used as a chromosomal DNA donor. Chromosomal DNA was prepared from the ATCC 13869 strain in accordance with an ordinary method. A DNA fragment containing dapB was amplified from the chromosomal DNA in accordance with PCR. As for DNA primers used for amplification, DNA's of 23-mers having nucleotide sequences shown in SEQ ID NOs: 17 and 18 in Sequence Listing respectively were synthesized in order to amplify a region of about 2.0 kb coding for DDPR on the basis of a sequence known for Brevibacterium lactofermentum (see Journal of Bacteriology, 157(9), 2743-2749 (1993)). Synthesis of DNA and PCR were performed in the same manner as described in Example 1. pCR-Script (produced by Invitrogen) was used as a cloning vector for the amplified gene fragment of 2,001 bp, and was ligated with the amplified dapB fragment. Thus a plasmid was constructed, in which the dapB fragment of 2,001 bp amplified from chromosome of Brevibacterium lactofermentum was ligated with pCR-Script. The plasmid obtained as described above, which had dapB originating from ATCC 13869, was designated as pCRDAPB. A transformant strain AJ13107 obtained by introducing pCRDAPB into E. coli JM109 strain has been internationally deposited since May 26, 1995 under an accession number of FERM BP-5114 in National Institute of Bioscience and Human Technology of Agency of Industrial Science and Technology of Ministry of International Trade and Industry (postal code: 305, 1-3, Higashi 1-chome, Tsukuba-shi, Ibaraki-ken, Japan) based on the Budapest Treaty.

A fragment of 1,101 bp containing a structural gene of DDPR was extracted by digesting pCRDAPB with EcoRV and SphI. This fragment was ligated with pHSG399 having been digested with HincII and SphI to prepare a plasmid. The prepared plasmid was designated as p399DPR.

Brevi.-ori was introduced into the prepared p399DPR to construct a plasmid carrying dapB autonomously replicable in coryneform bacteria. pHK4 was digested with a restriction enzyme KpnI (produced by Takara Shuzo), and cleaved ends were blunted. Blunt end formation was performed by using DNA Blunting kit (produced by Takara Shuzo) in accordance with a designated method. After the blunt end formation, a phosphorylated BamHI linker (produced by Takara Shuzo) was ligated to make modification so that the DNA fragment corresponding to the Brevi.-ori portion might be excised from pHK4 by digestion with only BamHI. This plasmid was digested with BamHI, and the generated Brevi.-ori DNA fragment was ligated with p399DPR having been also digested with BamHI to prepare a plasmid containing dapB autonomously replicable in coryneform bacteria. The prepared plasmid was designated as pDPRB. The process of construction of pDPRB is shown in Fig. 7.

(2) Determination of nucleotide sequence of dapB from Brevibacterium lactofermentum

Plasmid DNA was prepared from the AJ13107 strain harboring p399DPR, and its nucleotide sequence was determined in the same manner as described in Example 1. A determined nucleotide sequence and an amino acid sequence deduced from the nucleotide sequence are shown in SEQ ID NO: 19. Only the amino acid sequence is shown in SEQ ID NO: 20.

(3) Preparation of dapA from Brevibacterium lactofermentum

(1) Preparation of dapA and construction of plasmid carrying dapA

A wild type strain of Brevibacterium lactofermentum ATCC 13869 was used as a chromosomal DNA donor. Chro-

mosomal DNA was prepared from the ATCC 13869 strain in accordance with an ordinary method. A DNA fragment containing dapA was amplified from the chromosomal DNA in accordance with PCR. As for DNA primers used for amplification, DNA's of 23-mers having nucleotide sequences shown in SEQ ID NOs: 21 and 22 in Sequence Listing respectively were synthesized in rder to amplify a region of about 1.5 kb coding f r DDPS on th basis of a sequence known for Corynebacterium glutamicum (see Nucleic Acids Research, 18(21), 6421 (1990); EMBL accession No. X53993). Synthesis of DNA and PCR were performed in the same manner as described in Example 1. pCR1000 (produced by Invitrogen, see Bio/Technology, 9, 657-663 (1991)) was used as a cloning vector for the amplified gene fragment of 1,411 bp, and was ligated with the amplified dapA fragment. Ligation of DNA was performed by using DNA ligation kit (produced by Takara Shuzo) in accordance with a designated method. Thus a plasmid was constructed, in which the dapA fragment of 1,411 bp amplified from chromosome of Brevibacterium lactofermentum was ligated with pCR1000. The plasmid obtained as described above, which had dapA originating from ATCC 13869, was designated as pCRDAPA.

A transformant strain AJ13106 obtained by introducing pCRDAPA into E. coli JM109 strain has been internationally deposited since May 26, 1995 under an accession number of FERM BP-5113 in National Institute of Bioscience and Human Technology of Agency of Industrial Science and Technology of Ministry of International Trade and Industry (postal code: 305, 1-3, Higashi 1-chome, Tsukuba-shi, Ibaraki-ken, Japan) based on the Budapest Treaty.

Brevi.-ori was introduced into the prepared pCRDAPA to construct a plasmid carrying dapA autonomously replicable in coryneform bacteria. pHK4 was digested with restriction enzymes KpnI and BamHI (produced by Takara Shuzo), and cleaved ends were blunted. Blunt end formation was performed by using DNA Blunting kit (produced by Takara Shuzo) in accordance with a designated method. After the blunt end formation, a phosphorylated SmaI linker (produced by Takara Shuzo) was ligated to make modification so that the DNA fragment corresponding to the Brevi.-ori portion might be excised from pHK4 by digestion with only SmaI. This plasmid was digested with SmaI, and the generated Brevi.-ori DNA fragment was ligated with pCRDAPA having been also digested with SmaI to prepare a plasmid carrying dapA autonomously replicable in coryneform bacteria. This plasmid was designated as pDPSB. The process of construction of pDPSB(Km^r) is shown in Fig. 8.

(2) Determination of nucleotide sequence of dapA from Brevibacterium lactofermentum

Plasmid DNA was prepared from the AJ13106 strain harboring pCRDAPA, and its nucleotide sequence was determined in the same manner as described in Example 1. A determined nucleotide sequence and an amino acid sequence deduced from the nucleotide sequence are shown in SEQ ID NO: 23. Only the amino acid sequence is shown in SEQ ID NO: 24.

(4) Construction of Plasmid Carrying Both of Mutant lysC and dapA

A plasmid carrying mutant lysC, dapA, and replication origin of coryneform bacteria was constructed from the plasmid pCRDAPA carrying dapA and the plasmid p399AK9B carrying mutant lysC and Brevi.-ori. p399AK9B was completely digested with SalI, and then it was blunt-ended. An EcoRI linker was ligated therewith to construct a plasmid in which the SalI site was modified into an EcoRI site. The obtained plasmid was designated as p399AK9BSE. The mutant lysC and Brevi.-ori were excised as one fragment by partially digesting p399AK9BSE with EcoRI. This fragment was ligated with pCRDAPA having been digested with EcoRI. An obtained plasmid was designated as pCRCAB. This plasmid is autonomously replicable in E. coli and coryneform bacteria, and it gives kanamycin resistance to a host, the plasmid carrying both of mutant lysC and dapA. The process of construction of pCRCAB is shown in Fig. 9.

(5) Construction of Plasmid Carrying Both of Mutant lysC and dapB

A plasmid carrying mutant lysC and dapB was constructed from the plasmid p399AK9 carrying mutant lysC and the plasmid p399DPR carrying dapB. A fragment of 1,101 bp containing a structural gene of DDPR was extracted by digesting p399DPR with EcoRV and SphI. This fragment was ligated with p399AK9 having been digested with SalI and then blunt-ended and having been further digested with SphI to construct a plasmid carrying both of mutant lysC and dapB. This plasmid was designated as p399AKDDPR.

Next, Brevi.-ori was introduced into the obtained p399AKDDPR. The plasmid pHK4 containing Brevi.-ori was digested with a restriction enzyme KpnI (produced by Takara Shuzo), and cleaved ends were blunted. Blunt end formation was performed by using DNA Blunting kit (produced by Takara Shuzo) in accordance with a designated method. After the blunt end formation, a phosphorylated BamHI linker (produced by Takara Shuzo) was ligated to make modification so that the DNA fragment corresponding to the Brevi.-ori portion might be excised from pHK4 by digestion with only BamHI. This plasmid was digested with BamHI, and the generated Brevi.-ori DNA fragment was ligated with p399AKDDPR having been also digested with BamHI to construct a plasmid carrying mutant lysC and dapB autonomously replicable in coryneform bacteria. The constructed plasmid was designated as pCB. The process of construc-

tion of pCB is shown in Fig. 10.

(6) Construction of Plasmid Carrying Both of dapA and dapB

The plasmid pCRDAPA carrying dapA was digested with KpnI and EcoRI to extract a DNA fragment containing dapA and the fragment was ligated with the vector plasmid pHSG399 having been digested with KpnI and EcoRI. An obtained plasmid was designated as p399DPS.

On the other hand, the plasmid pCRDAPB carrying dapB was digested with SacII and EcoRI to extract a DNA fragment of 2.0 kb containing a region coding for DDPR and the fragment was ligated with p399DPS having been digested with SacII and EcoRI to construct a plasmid carrying both of dapA and dapB. The obtained plasmid was designated as p399AB.

Next, Brevi.-ori was introduced into p399AB. pHK4 carrying Brevi.-ori was digested with a restriction enzyme BamHI (produced by Takara Shuzo), and cleaved ends were blunted. Blunt end formation was performed by using DNA Blunting kit (produced by Takara Shuzo) in accordance with a designated method. After the blunt end formation, a phosphorylated KpnI linker (produced by Takara Shuzo) was ligated to make modification so that the DNA fragment corresponding to the Brevi.-ori portion might be excised from pHK4 by digestion with only KpnI. This plasmid was digested with KpnI, and the generated Brevi.-ori DNA fragment was ligated with p399AB having been also digested with KpnI to construct a plasmid carrying dapA and dapB autonomously replicable in coryneform bacteria. The constructed plasmid was designated as pAB. The process of construction of pAB is shown in Fig. 11.

(7) Construction of Plasmid Carrying Mutant lysC, dapA, and dapB Together

p399DPS was digested with EcoRI and SphI followed by blunt end formation to extract a dapA gene fragment. This fragment was ligated with the p399AK9 having been digested with SalI and blunt-ended to construct a plasmid p399CA in which mutant lysC and dapA co-existed.

The plasmid pCRDAPB carrying dapB was digested with EcoRI and blunt-ended, followed by digestion with SacI to extract a DNA fragment of 2.0 kb comprising dapB. The plasmid p399CA carrying dapA and mutant lysC was digested with SpeI and blunt-ended, and was thereafter digested with SacI and ligated with the extracted dapB fragment to obtain a plasmid carrying mutant lysC, dapA, and dapB. This plasmid was designated as p399CAB.

Next, Brevi.-ori was introduced into p399CAB. The plasmid pHK4 carrying Brevi.-ori was digested with a restriction enzyme BamHI (produced by Takara Shuzo), and cleaved ends were blunted. Blunt end formation was performed by using DNA Blunting kit (produced by Takara Shuzo) in accordance with a designated method. After the blunt end formation, a phosphorylated KpnI linker (produced by Takara Shuzo) was ligated to make modification so that the DNA fragment corresponding to the Brevi.-ori portion might be excised from pHK4 by digestion with only KpnI. This plasmid was digested with KpnI, and the generated Brevi.-ori DNA fragment was ligated with p399CAB having been also digested with KpnI to construct a plasmid carrying mutant lysC, dapA, and dapB together autonomously replicable in coryneform bacteria. The constructed plasmid was designated as pCAB. The process of construction of pCAB is shown in Fig. 12.

(8) Construction of Plasmid Carrying Mutant lysC, dapA, dapB, and lysA Together

The plasmid p299LYSA carrying lysA was digested with KpnI and BamHI and blunt-ended, and then a lysA gene fragment was extracted. This fragment was ligated with pCAB having been digested with HpaI (produced by Takara Shuzo) and blunt-ended to construct a plasmid carrying mutant lysC, dapA, dapB, and lysA together autonomously replicable in coryneform bacteria. The constructed plasmid was designated as pCABL. The process of construction of pCABL is shown in Fig. 13. It is noted that the lysA gene fragment is inserted into a HpaI site in a DNA fragment containing the dapB gene in pCABL, however, the HpaI site is located upstream from a promoter for the dapB gene (nucleotide numbers 611 to 616 in SEQ ID NO: 19), and the dapB gene is not divided.

(9) Construction of Plasmid Carrying Mutant lysC, dapA, dapB, ddh, and lysA Together

pHSG299 was digested with XbaI and KpnI, and was ligated with p399DL carrying ddh and lysA having been digested with XbaI and KpnI. A constructed plasmid was designated as p299DL. p299DL was digested with XbaI and KpnI and blunt-ended. After the blunt end formation, a DNA fragment carrying ddh and lysA was extracted. This DNA fragment was ligated with the plasmid pCAB carrying mutant lysC, dapA, and dapB together having been digested with HpaI and blunt-ended to construct a plasmid carrying mutant lysC, dapA, dapB, lysA and ddh together autonomously replicable in coryneform bacteria. The constructed plasmid was designated as pCABDL. The process of construction of pCABDL is shown in Fig. 14.

Example 5: Introduction of Plasmids Carrying Genes for L-Lysine Biosynthesis into L-Lysine-Producing Bacterium of Brevibacterium lactofermentum

The plasmids carrying the genes for L-lysine biosynthesis constructed as described above, namely pLYSAB(Cm^r), pPK4D(Cm^r), p399AK9B(Cm^r), pDPSB(Km^r), pDPRB(Cm^r), pCRCAB(Km^r), pAB(Cm^r), pDL(Cm^r), pCB(Cm^r), pCAB(Cm^r), pCABL(Cm^r), and pCABDL(Cm^r) were introduced into an L-lysine-producing bacterium AJ11082 (NRRL B-11470) of Brevibacterium lactofermentum respectively. AJ11082 strain has a property of AEC resistance. The plasmids were introduced in accordance with an electric pulse method (Sugimoto et al., Japanese Patent Application Laid-open No. 2-207791). Transformants were selected based on drug resistance markers possessed by the respective plasmids. Transformants were selected on a complete medium containing 5 μg/ml of chloramphenicol when a plasmid carrying a chloramphenicol resistance gene was introduced, or transformants were selected on a complete medium containing 25 μg/ml of kanamycin when a plasmid carrying a kanamycin resistance gene was introduced.

Example 6: Production of L-Lysine

Each of the transformants obtained in Example 5 was cultivated in an L-lysine-producing medium to evaluate its L-lysine productivity. The L-lysine-producing medium had the following composition.

[L-Lysine-producing medium]

The following components other than calcium carbonate (per liter) were dissolved to make adjustment at pH 8.0 with KOH. The medium was sterilized at 115°C for 15 minutes, and calcium carbonate (50 g) having been separately sterilized in hot air in a dry state was added to the sterilized medium.

Glucose	100 g
$(NH_4)_2SO_4$	55 g
KH_2PO_4	1 g
$MgSO_4 \cdot 7H_2O$	1 g
Biotin	500 μg
Thiamin	2000 μg
$FeSO_4 \cdot 7H_2O$	0.01 g
$MnSO_4 \cdot 7H_2O$	0.01 g
Nicotinamide	5 mg
Protein hydrolysate (Mamenou)	30 ml
Calcium carbonate	50 g

Each of the various types of the transformants and the parent strain was inoculated to the medium having the composition described above to perform cultivation at 31.5°C with reciprocating shaking. The amount of produced L-lysine after 40 or 72 hours of cultivation, and the growth after 72 hours (OD_{562}) are shown in Table 1. In the table, lysC* represents mutant lysC. The growth was quantitatively determined by measuring OD at 560 nm after 101-fold dilution.

Table 1

Accumulation of L-Lysine after Cultivation for 40 or 72 Hours				
Bacterial strain /plasmid	Introduced gene	Amount of produced L-lysine(g/L)		Growth (OD_{562}/101)
		after 40 hrs	after 72 hrs	
AJ11082		22.0	29.8	0.450
AJ11082/pLYSAB	lysA	19.8	32.5	0.356
AJ11082/pPK4D	ddh	19.0	33.4	0.330
AJ11082/p399AK9B	lysC*	16.8	34.5	0.398
AJ11082/pDPSB	dapA	18.7	33.8	0.410
AJ11082/pDPRB	dapB	19.9	29.9	0.445
AJ11082/pCRCAB	lysC*, dapA	19.7	36.5	0.360
AJ11082/pAB	dapA, dapB	19.0	34.8	0.390
AJ11082/pDL	lysA, ddh	23.3	31.6	0.440
AJ11082/pCB	lysC*, dapB	23.3	35.0	0.440
AJ11082/pCAB	lysC*, dapA, dapB	23.0	45.0	0.425
AJ11082/pCABL	lysC*, dapA, dapB, lysA	26.2	46.5	0.379
AJ11082/pCABDL	lysC*, dapA, dapB, lysA, ddh	26.5	47.0	0.409

As shown above, when lysA, ddh, mutant lysC, dapA, or dapB was enhanced singly, the amount of produced L-lysine after 72 hours of cultivation was larger than or equivalent to that produced by the parent strain, however, the amount of produced L-lysine after 40 hours of cultivation was smaller than that produced by the parent strain. Namely, the L-lysine-producing speed was lowered in cultivation for a short period. Similarly, when mutant lysC and dapA, or dapA and dapB were enhanced in combination, the amount of produced L-lysine after 72 hours of cultivation was larger than that produced by the parent strain, however, the amount of produced L-lysine after 40 hours of cultivation was smaller than that produced by the parent strain. Thus the L-lysine-producing speed was lowered.

On the other hand, when only lysA and ddh were enhanced in combination, the growth was improved, the L-lysine-producing speed was successfully restored in the short period of cultivation, and the accumulated amount of L-lysine was also improved in cultivation for a long period.

Also, in the case of the strain in which dapB was enhanced together with mutant lysC, and in the case of the strain in which dapA as well as these genes were simultaneously enhanced, the growth was improved and the L-lysine-producing speed was increased compared with the parent strain. In the case of the strain in which these three genes were simultaneously enhanced, both of the L-lysine-producing speed and the amount of accumulated L-lysine were further improved by further enhancing lysA and ddh.

SEQUENCE LISTING

(1) GENERAL INFORMATION:

(i) APPLICANT: AJINOMOTO CO., LTD.

(ii) TITLE OF INVENTION: METHOD OF PRODUCING L-LYSINE

(iii) NUMBER OF SEQUENCES: 24

(iv) CORRESPONDENCE ADDRESS:

(A) ADDRESSEE:

(B) STREET:

(C) CITY:

(E) COUNTRY:

(F) ZIP:

(v) COMPUTER READABLE FORM:

(A) MEDIUM TYPE: Floppy disk

(B) COMPUTER: IBM PC compatible

(C) OPERATING SYSTEM: PC-DOS/MS-DOS

(D) SOFTWARE: PatentIn Release #1.0, Version #1.30

(vi) CURRENT APPLICATION DATA:

(A) APPLICATION NUMBER:

(B) FILING DATE:

(C) CLASSIFICATION:

(vii) PRIOR APPLICATION DATA:

(A) APPLICATION NUMBER: JP 8-142812

(B) FILING DATE: 05-JUN-1996

(viii) ATTORNEY/AGENT INFORMATION:

(A) NAME:

(B) REGISTRATION NUMBER:

(ix) TELECOMMUNICATION INFORMATION:

(A) TELEPHONE:

(B) TELEFAX:

(2) INFORMATION FOR SEQ ID NO: 1

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 23 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: other nucleic acid

(A) DESCRIPTION: /desc="Synthetic DNA"

(iv) ANTI-SENSE: NO

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:1:

GTGGAGCCGA CCATTCCGCG AGG 23

(2) INFORMATION FOR SEQ ID NO: 2

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 23 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: other nucleic acid

(A) DESCRIPTION: /desc="Synthetic DNA"

(iv) ANTI-SENSE: YES

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:

CCAAAACCGC CCTCCACGGC GAA 23

(2) INFORMATION FOR SEQ ID NO: 3

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 3579 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: double

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: Genomic DNA

(vi) ORIGINAL SOURCE:

(A) ORGANISM: Brevibacterium lactofermentum

(B) STRAIN: ATCC 13869

(ix) FEATURE:

(A) NAME/KEY: CDS

(B) LOCATION: 533..2182

(ix) FEATURE:

(A) NAME/KEY: CDS

(B) LOCATION: 2188..3522

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:3:

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GTTACCGAAG ATGGTGCCGT GCTTTTCGCC TTGGGCAGGG ACCTTGACAA AGCCCACGCT     180
GATATCGCCA AGTGAGGGAT CAGAATAGTG CATGGGCACG TCGATGCTGC CACATTGAGC     240
GGAGGCAATA TCTACCTGAG GTGGGCATTC TTCCCAGCGG ATGTTTTCTT GCGCTGCTGC     300
AGTGGGCATT GATACCAAAA AGGGGCTAAG CGCAGTCGAG GCGGCAAGAA CTGCTACTAC     360
CCTTTTTATT GTCGAACGGG GCATTACGGC TCCAAGGACG TTTGTTTTCT GGGTCAGTTA     420
CCCCAAAAAG CATATACAGA GACCAATGAT TTTTCATTAA AAAGGCAGGG ATTTGTTATA     480
AGTATGGGTC GTATTCTGTG CGACGGGTGT ACCTCGGCTA GAATTTCTCC CC ATG         535
                                                          Met
                                                           1
ACA CCA GCT GAT CTC GCA ACA TTG ATT AAA GAG ACC GCG GTA GAG GTT       583
Thr Pro Ala Asp Leu Ala Thr Leu Ile Lys Glu Thr Ala Val Glu Val
                5                  10                  15
TTG ACC TCC CGC GAG CTC GAT ACT TCT GTT CTT CCG GAG CAG GTA GTT       631
Leu Thr Ser Arg Glu Leu Asp Thr Ser Val Leu Pro Glu Gln Val Val
            20                  25                  30
GTG GAG CGT CCG CGT AAC CCA GAG CAC GGC GAT TAC GCC ACC AAC ATT       679
Val Glu Arg Pro Arg Asn Pro Glu His Gly Asp Tyr Ala Thr Asn Ile
        35                  40                  45
GCA TTG CAG GTG GCT AAA AAG GTC GGT CAG AAC CCT CGG GAT TTG GCT       727
Ala Leu Gln Val Ala Lys Lys Val Gly Gln Asn Pro Arg Asp Leu Ala
    50                  55                  60                  65
ACC TGG CTG GCA GAG GCA TTG GCT GCA GAT GAC GCC ATT GAT TCT GCT       775
Thr Trp Leu Ala Glu Ala Leu Ala Ala Asp Asp Ala Ile Asp Ser Ala
                70                  75                  80
GAA ATT GCT GGC CCA GGC TTT TTG AAC ATT CGC CTT GCT GCA GCA GCA       823
Glu Ile Ala Gly Pro Gly Phe Leu Asn Ile Arg Leu Ala Ala Ala Ala
            85                  90                  95
CAG GGT GAA ATT GTG GCC AAG ATT CTG GCA CAG GGC GAG ACT TTC GGA       871
Gln Gly Glu Ile Val Ala Lys Ile Leu Ala Gln Gly Glu Thr Phe Gly
        100                 105                 110
AAC TCC GAT CAC CTT TCC CAC TTG GAC GTG AAC CTC GAG TTC GTT TCT       919
Asn Ser Asp His Leu Ser His Leu Asp Val Asn Leu Glu Phe Val Ser
    115                 120                 125
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GCA AAC CCA ACC GGA CCT ATT CAC CTT GGC GGA ACC CGC TGG GCT GCC      967
Ala Asn Pro Thr Gly Pro Ile His Leu Gly Gly Thr Arg Trp Ala Ala
130                 135                 140                 145
GTG GGT GAC TCT TTG GGT CGT GTG CTG GAG GCT TCC GGC GCG AAA GTG     1015
Val Gly Asp Ser Leu Gly Arg Val Leu Glu Ala Ser Gly Ala Lys Val
                150                 155                 160
ACC CGC GAA TAC TAC TTC AAC GAT CAC GGT CGC CAG ATC GAT CGT TTC     1063
Thr Arg Glu Tyr Tyr Phe Asn Asp His Gly Arg Gln Ile Asp Arg Phe
            165                 170                 175
GCT TTG TCC CTT CTT GCA GCG GCG AAG GGC GAG CCA ACG CCA GAA GAC     1111
Ala Leu Ser Leu Leu Ala Ala Ala Lys Gly Glu Pro Thr Pro Glu Asp
        180                 185                 190
GGT TAT GGC GGC GAA TAC ATT AAG GAA ATT GCG GAG GCA ATC GTC GAA     1159
Gly Tyr Gly Gly Glu Tyr Ile Lys Glu Ile Ala Glu Ala Ile Val Glu
    195                 200                 205
AAG CAT CCT GAA GCG TTG GCT TTG GAG CCT GCC GCA ACC CAG GAG CTT     1207
Lys His Pro Glu Ala Leu Ala Leu Glu Pro Ala Ala Thr Gln Glu Leu
210                 215                 220                 225
TTC CGC GCT GAA GGC GTG GAG ATG ATG TTC GAG CAC ATC AAA TCT TCC     1255
Phe Arg Ala Glu Gly Val Glu Met Met Phe Glu His Ile Lys Ser Ser
                230                 235                 240
CTG CAT GAG TTC GGC ACC GAT TTC GAT GTC TAC TAC CAC GAG AAC TCC     1303
Leu His Glu Phe Gly Thr Asp Phe Asp Val Tyr Tyr His Glu Asn Ser
            245                 250                 255
CTG TTC GAG TCC GGT GCG GTG GAC AAG GCC GTG CAG GTG CTG AAG GAC     1351
Leu Phe Glu Ser Gly Ala Val Asp Lys Ala Val Gln Val Leu Lys Asp
        260                 265                 270
AAC GGC AAC CTG TAC GAA AAC GAG GGC GCT TGG TGG CTG CGT TCC ACC     1399
Asn Gly Asn Leu Tyr Glu Asn Glu Gly Ala Trp Trp Leu Arg Ser Thr
    275                 280                 285
GAA TTC GGC GAT GAC AAA GAC CGC GTG GTG ATC AAG TCT GAC GGC GAC     1447
Glu Phe Gly Asp Asp Lys Asp Arg Val Val Ile Lys Ser Asp Gly Asp
290                 295                 300                 305
GCA GCC TAC ATC GCT GGC GAT ATC GCG TAC GTG GCT GAT AAG TTC TCC     1495
Ala Ala Tyr Ile Ala Gly Asp Ile Ala Tyr Val Ala Asp Lys Phe Ser
                310                 315                 320
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CGC GGA CAC AAC CTA AAC ATC TAC ATG TTG GGT GCT GAC CAC CAT GGT      1543
Arg Gly His Asn Leu Asn Ile Tyr Met Leu Gly Ala Asp His His Gly
            325                 330                 335
TAC ATC GCG CGC CTG AAG GCA GCG GCG GCG GCA CTT GGC TAC AAG CCA      1591
Tyr Ile Ala Arg Leu Lys Ala Ala Ala Ala Ala Leu Gly Tyr Lys Pro
        340                 345                 350
GAA GGC GTT GAA GTC CTG ATT GGC CAG ATG GTG AAC CTG CTT CGC GAC      1639
Glu Gly Val Glu Val Leu Ile Gly Gln Met Val Asn Leu Leu Arg Asp
    355                 360                 365
GGC AAG GCA GTG CGT ATG TCC AAG CGT GCA GGC ACC GTG GTC ACC CTA      1687
Gly Lys Ala Val Arg Met Ser Lys Arg Ala Gly Thr Val Val Thr Leu
370                 375                 380                 385
GAT GAC CTC GTT GAA GCA ATC GGC ATC GAT GCG GCG CGT TAC TCC CTG      1735
Asp Asp Leu Val Glu Ala Ile Gly Ile Asp Ala Ala Arg Tyr Ser Leu
                390                 395                 400
ATC CGT TCC TCC GTG GAT TCT TCC CTG GAT ATC GAT CTC GGC CTG TGG      1783
Ile Arg Ser Ser Val Asp Ser Ser Leu Asp Ile Asp Leu Gly Leu Trp
            405                 410                 415
GAA TCC CAG TCC TCC GAC AAC CCT GTG TAC TAC GTG CAG TAC GGA CAC      1831
Glu Ser Gln Ser Ser Asp Asn Pro Val Tyr Tyr Val Gln Tyr Gly His
        420                 425                 430
GCT CGT CTG TGC TCC ATC GCG CGC AAG GCA GAG ACC TTG GGT GTC ACC      1879
Ala Arg Leu Cys Ser Ile Ala Arg Lys Ala Glu Thr Leu Gly Val Thr
    435                 440                 445
GAG GAA GGC GCA GAC CTA TCT CTA CTG ACC CAC GAC CGC GAA GGC GAT      1927
Glu Glu Gly Ala Asp Leu Ser Leu Leu Thr His Asp Arg Glu Gly Asp
450                 455                 460                 465
CTC ATC CGC ACA CTC GGA GAG TTC CCA GCA GTG GTG AAG GCT GCC GCT      1975
Leu Ile Arg Thr Leu Gly Glu Phe Pro Ala Val Val Lys Ala Ala Ala
                470                 475                 480
GAC CTA CGT GAA CCA CAC CGC ATT GCC CGC TAT GCT GAG GAA TTA GCT      2023
Asp Leu Arg Glu Pro His Arg Ile Ala Arg Tyr Ala Glu Glu Leu Ala
            485                 490                 495
GGA ACT TTC CAC CGC TTC TAC GAT TCC TGC CAC ATC CTT CCA AAG GTT      2071
Gly Thr Phe His Arg Phe Tyr Asp Ser Cys His Ile Leu Pro Lys Val
        500                 505                 510
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GAT GAG GAT ACG GCA CCA ATC CAC ACA GCA CGT CTG GCA CTT GCA GCA     2119
Asp Glu Asp Thr Ala Pro Ile His Thr Ala Arg Leu Ala Leu Ala Ala
    515                 520                 525
GCA ACC CGC CAG ACC CTC GCT AAC GCC CTG CAC CTG GTT GGC GTT TCC     2167
Ala Thr Arg Gln Thr Leu Ala Asn Ala Leu His Leu Val Gly Val Ser
530                 535                 540                 545
GCA CCG GAG AAG ATG TAACA ATG GCT ACA GTT GAA AAT TTC AAT GAA       2214
Ala Pro Glu Lys Met       Met Ala Thr Val Glu Asn Phe Asn Glu
                550         1               5
CTT CCC GCA CAC GTA TGG CCA CGC AAT GCC GTG CGC CAA GAA GAC GGC     2262
Leu Pro Ala His Val Trp Pro Arg Asn Ala Val Arg Gln Glu Asp Gly
 10                  15                  20                  25
GTT GTC ACC GTC GCT GGT GTG CCT CTG CCT GAC CTC GCT GAA GAA TAC     2310
Val Val Thr Val Ala Gly Val Pro Leu Pro Asp Leu Ala Glu Glu Tyr
                30                  35                  40
GGA ACC CCA CTG TTC GTA GTC GAC GAG GAC GAT TTC CGT TCC CGC TGT     2358
Gly Thr Pro Leu Phe Val Val Asp Glu Asp Asp Phe Arg Ser Arg Cys
            45                  50                  55
CGC GAC ATG GCT ACC GCA TTC GGT GGA CCA GGC AAT GTG CAC TAC GCA     2406
Arg Asp Met Ala Thr Ala Phe Gly Gly Pro Gly Asn Val His Tyr Ala
        60                  65                  70
TCT AAA GCG TTC CTG ACC AAG ACC ATT GCA CGT TGG GTT GAT GAA GAG     2454
Ser Lys Ala Phe Leu Thr Lys Thr Ile Ala Arg Trp Val Asp Glu Glu
    75                  80                  85
GGG CTG GCA CTG GAC ATT GCA TCC ATC AAC GAA CTG GGC ATT GCC CTG     2502
Gly Leu Ala Leu Asp Ile Ala Ser Ile Asn Glu Leu Gly Ile Ala Leu
 90                  95                  100                 105
GCC GCT GGT TTC CCC GCC AGC CGT ATC ACC GCG CAC GGC AAC AAC AAA     2550
Ala Ala Gly Phe Pro Ala Ser Arg Ile Thr Ala His Gly Asn Asn Lys
                110                 115                 120
GGC GTA GAG TTC CTG CGC GCG TTG GTT CAA AAC GGT GTG GGA CAC GTG     2598
Gly Val Glu Phe Leu Arg Ala Leu Val Gln Asn Gly Val Gly His Val
            125                 130                 135
GTG CTG GAC TCC GCA CAG GAA CTA GAA CTG TTG GAT TAC GTT GCC GCT     2646
Val Leu Asp Ser Ala Gln Glu Leu Glu Leu Leu Asp Tyr Val Ala Ala
        140                 145                 150
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GGT GAA GGC AAG ATT CAG GAC GTG TTG ATC CGC GTA AAG CCA GGC ATC      2694
Gly Glu Gly Lys Ile Gln Asp Val Leu Ile Arg Val Lys Pro Gly Ile
    155                 160                 165
GAA GCA CAC ACC CAC GAG TTC ATC GCC ACT AGC CAC GAA GAC CAG AAG      2742
Glu Ala His Thr His Glu Phe Ile Ala Thr Ser His Glu Asp Gln Lys
170                 175                 180                 185
TTC GGA TTC TCC CTG GCA TCC GGT TCC GCA TTC GAA GCA GCA AAA GCC      2790
Phe Gly Phe Ser Leu Ala Ser Gly Ser Ala Phe Glu Ala Ala Lys Ala
                190                 195                 200
GCC AAC AAC GCA GAA AAC CTG AAC CTG GTT GGC CTG CAC TGC CAC GTT      2838
Ala Asn Asn Ala Glu Asn Leu Asn Leu Val Gly Leu His Cys His Val
            205                 210                 215
GGT TCC CAG GTG TTC GAC GCC GAA GGC TTC AAG CTG GCA GCA GAA CGC      2886
Gly Ser Gln Val Phe Asp Ala Glu Gly Phe Lys Leu Ala Ala Glu Arg
        220                 225                 230
GTG TTG GGC CTG TAC TCA CAG ATC CAC AGC GAA CTG GGC GTT GCC CTT      2934
Val Leu Gly Leu Tyr Ser Gln Ile His Ser Glu Leu Gly Val Ala Leu
    235                 240                 245
CCT GAA CTG GAT CTC GGT GGC GGA TAC GGC ATT GCC TAT ACC GCA GCT      2982
Pro Glu Leu Asp Leu Gly Gly Gly Tyr Gly Ile Ala Tyr Thr Ala Ala
250                 255                 260                 265
GAA GAA CCA CTC AAC GTC GCA GAA GTT GCC TCC GAC CTG CTC ACC GCA      3030
Glu Glu Pro Leu Asn Val Ala Glu Val Ala Ser Asp Leu Leu Thr Ala
                270                 275                 280
GTC GGA AAA ATG GCA GCG GAA CTA GGC ATC GAC GCA CCA ACC GTG CTT      3078
Val Gly Lys Met Ala Ala Glu Leu Gly Ile Asp Ala Pro Thr Val Leu
            285                 290                 295
GTT GAG CCC GGC CGC GCT ATC GCA GGC CCC TCC ACC GTG ACC ATC TAC      3126
Val Glu Pro Gly Arg Ala Ile Ala Gly Pro Ser Thr Val Thr Ile Tyr
        300                 305                 310
GAA GTC GGC ACC ACC AAA GAC GTC CAC GTA GAC GAC GAC AAA ACC CGC      3174
Glu Val Gly Thr Thr Lys Asp Val His Val Asp Asp Asp Lys Thr Arg
    315                 320                 325
CGT TAC ATC GCC GTG GAC GGA GGC ATG TCC GAC AAC ATC CGC CCA GCA      3222
Arg Tyr Ile Ala Val Asp Gly Gly Met Ser Asp Asn Ile Arg Pro Ala
330                 335                 340                 345
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CTC TAC GGC TCC GAA TAC GAC GCC CGC GTA GTA TCC CGC TTC GCC GAA      3270
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                350                 355                 360
GGA GAC CCA GTA AGC ACC CGC ATC GTG GGC TCC CAC TGC GAA TCC GGC      3318
Gly Asp Pro Val Ser Thr Arg Ile Val Gly Ser His Cys Glu Ser Gly
            365                 370                 375
GAT ATC CTG ATC AAC GAT GAA ATC TAC CCA TCT GAC ATC ACC AGC GGC      3366
Asp Ile Leu Ile Asn Asp Glu Ile Tyr Pro Ser Asp Ile Thr Ser Gly
        380                 385                 390
GAC TTC CTT GCA CTC GCA GCC ACC GGC GCA TAC TGC TAC GCC ATG AGC      3414
Asp Phe Leu Ala Leu Ala Ala Thr Gly Ala Tyr Cys Tyr Ala Met Ser
    395                 400                 405
TCC CGC TAC AAC GCC TTC ACA CGG CCC GCC GTC GTG TCC GTC CGC GCT      3462
Ser Arg Tyr Asn Ala Phe Thr Arg Pro Ala Val Val Ser Val Arg Ala
410                 415                 420                 425
GGC AGC TCC CGC CTC ATG CTG CGC CGC GAA ACG CTC GAC GAC ATC CTC      3510
Gly Ser Ser Arg Leu Met Leu Arg Arg Glu Thr Leu Asp Asp Ile Leu
                430                 435                 440
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            445
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(2) INFORMATION FOR SEQ ID NO: 4

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 550 amino acids

(B) TYPE: amino acid

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:4:

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Val Leu Thr Ser Arg Glu Leu Asp Thr Ser Val Leu Pro Glu Gln Val
            20                  25                  30
Val Val Glu Arg Pro Arg Asn Pro Glu His Gly Asp Tyr Ala Thr Asn
        35                  40                  45
```

Ile Ala Leu Gln Val Ala Lys Lys Val Gly Gln Asn Pro Arg Asp Leu
50 55 60
Ala Thr Trp Leu Ala Glu Ala Leu Ala Ala Asp Asp Ala Ile Asp Ser
65 70 75 80
Ala Glu Ile Ala Gly Pro Gly Phe Leu Asn Ile Arg Leu Ala Ala Ala
85 90 95
Ala Gln Gly Glu Ile Val Ala Lys Ile Leu Ala Gln Gly Glu Thr Phe
100 105 110
Gly Asn Ser Asp His Leu Ser His Leu Asp Val Asn Leu Glu Phe Val
115 120 125
Ser Ala Asn Pro Thr Gly Pro Ile His Leu Gly Gly Thr Arg Trp Ala
130 135 140
Ala Val Gly Asp Ser Leu Gly Arg Val Leu Glu Ala Ser Gly Ala Lys
145 150 155 160
Val Thr Arg Glu Tyr Tyr Phe Asn Asp His Gly Arg Gln Ile Asp Arg
165 170 175
Phe Ala Leu Ser Leu Leu Ala Ala Ala Lys Gly Glu Pro Thr Pro Glu
180 185 190
Asp Gly Tyr Gly Gly Glu Tyr Ile Lys Glu Ile Ala Glu Ala Ile Val
195 200 205
Glu Lys His Pro Glu Ala Leu Ala Leu Glu Pro Ala Ala Thr Gln Glu
210 215 220
Leu Phe Arg Ala Glu Gly Val Glu Met Met Phe Glu His Ile Lys Ser
225 230 235 240
Ser Leu His Glu Phe Gly Thr Asp Phe Asp Val Tyr Tyr His Glu Asn
245 250 255
Ser Leu Phe Glu Ser Gly Ala Val Asp Lys Ala Val Gln Val Leu Lys
260 265 270
Asp Asn Gly Asn Leu Tyr Glu Asn Glu Gly Ala Trp Trp Leu Arg Ser
275 280 285
Thr Glu Phe Gly Asp Asp Lys Asp Arg Val Val Ile Lys Ser Asp Gly
290 295 300
Asp Ala Ala Tyr Ile Ala Gly Asp Ile Ala Tyr Val Ala Asp Lys Phe
305 310 315 320
Ser Arg Gly His Asn Leu Asn Ile Tyr Met Leu Gly Ala Asp His His
325 330 335

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Gly Tyr Ile Ala Arg Leu Lys Ala Ala Ala Ala Ala Leu Gly Tyr Lys
            340                 345                 350
Pro Glu Gly Val Glu Val Leu Ile Gly Gln Met Val Asn Leu Leu Arg
        355                 360                 365
Asp Gly Lys Ala Val Arg Met Ser Lys Arg Ala Gly Thr Val Val Thr
    370                 375                 380
Leu Asp Asp Leu Val Glu Ala Ile Gly Ile Asp Ala Ala Arg Tyr Ser
385                 390                 395                 400
Leu Ile Arg Ser Ser Val Asp Ser Ser Leu Asp Ile Asp Leu Gly Leu
                405                 410                 415
Trp Glu Ser Gln Ser Ser Asp Asn Pro Val Tyr Tyr Val Gln Tyr Gly
            420                 425                 430
His Ala Arg Leu Cys Ser Ile Ala Arg Lys Ala Glu Thr Leu Gly Val
        435                 440                 445
Thr Glu Glu Gly Ala Asp Leu Ser Leu Leu Thr His Asp Arg Glu Gly
    450                 455                 460
Asp Leu Ile Arg Thr Leu Gly Glu Phe Pro Ala Val Val Lys Ala Ala
465                 470                 475                 480
Ala Asp Leu Arg Glu Pro His Arg Ile Ala Arg Tyr Ala Glu Glu Leu
                485                 490                 495
Ala Gly Thr Phe His Arg Phe Tyr Asp Ser Cys His Ile Leu Pro Lys
            500                 505                 510
Val Asp Glu Asp Thr Ala Pro Ile His Thr Ala Arg Leu Ala Leu Ala
        515                 520                 525
Ala Ala Thr Arg Gln Thr Leu Ala Asn Ala Leu His Leu Val Gly Val
    530                 535                 540
Ser Ala Pro Glu Lys Met
545                 550
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(2) INFORMATION FOR SEQ ID NO: 5

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 445 amino acids

(B) TYPE: amino acid

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:5:

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Met Ala Thr Val Glu Asn Phe Asn Glu Leu Pro Ala His Val Trp Pro
 1               5                  10                  15
Arg Asn Ala Val Arg Gln Glu Asp Gly Val Val Thr Val Ala Gly Val
            20                  25                  30
Pro Leu Pro Asp Leu Ala Glu Glu Tyr Gly Thr Pro Leu Phe Val Val
        35                  40                  45
Asp Glu Asp Asp Phe Arg Ser Arg Cys Arg Asp Met Ala Thr Ala Phe
    50                  55                  60
Gly Gly Pro Gly Asn Val His Tyr Ala Ser Lys Ala Phe Leu Thr Lys
65                  70                  75                  80
Thr Ile Ala Arg Trp Val Asp Glu Glu Gly Leu Ala Leu Asp Ile Ala
                85                  90                  95
Ser Ile Asn Glu Leu Gly Ile Ala Leu Ala Ala Gly Phe Pro Ala Ser
            100                 105                 110
Arg Ile Thr Ala His Gly Asn Asn Lys Gly Val Glu Phe Leu Arg Ala
        115                 120                 125
Leu Val Gln Asn Gly Val Gly His Val Val Leu Asp Ser Ala Gln Glu
    130                 135                 140
Leu Glu Leu Leu Asp Tyr Val Ala Ala Gly Glu Gly Lys Ile Gln Asp
145                 150                 155                 160
Val Leu Ile Arg Val Lys Pro Gly Ile Glu Ala His Thr His Glu Phe
                165                 170                 175
Ile Ala Thr Ser His Glu Asp Gln Lys Phe Gly Phe Ser Leu Ala Ser
            180                 185                 190
Gly Ser Ala Phe Glu Ala Ala Lys Ala Ala Asn Asn Ala Glu Asn Leu
        195                 200                 205
Asn Leu Val Gly Leu His Cys His Val Gly Ser Gln Val Phe Asp Ala
    210                 215                 220
Glu Gly Phe Lys Leu Ala Ala Glu Arg Val Leu Gly Leu Tyr Ser Gln
225                 230                 235                 240
Ile His Ser Glu Leu Gly Val Ala Leu Pro Glu Leu Asp Leu Gly Gly
                245                 250                 255
Gly Tyr Gly Ile Ala Tyr Thr Ala Ala Glu Glu Pro Leu Asn Val Ala
            260                 265                 270
```

```
Glu Val Ala Ser Asp Leu Leu Thr Ala Val Gly Lys Met Ala Ala Glu
        275                 280                 285
Leu Gly Ile Asp Ala Pro Thr Val Leu Val Glu Pro Gly Arg Ala Ile
    290                 295                 300
Ala Gly Pro Ser Thr Val Thr Ile Tyr Glu Val Gly Thr Thr Lys Asp
305                 310                 315                 320
Val His Val Asp Asp Asp Lys Thr Arg Arg Tyr Ile Ala Val Asp Gly
                325                 330                 335
Gly Met Ser Asp Asn Ile Arg Pro Ala Leu Tyr Gly Ser Glu Tyr Asp
            340                 345                 350
Ala Arg Val Val Ser Arg Phe Ala Glu Gly Asp Pro Val Ser Thr Arg
        355                 360                 365
Ile Val Gly Ser His Cys Glu Ser Gly Asp Ile Leu Ile Asn Asp Glu
    370                 375                 380
Ile Tyr Pro Ser Asp Ile Thr Ser Gly Asp Phe Leu Ala Leu Ala Ala
385                 390                 395                 400
Thr Gly Ala Tyr Cys Tyr Ala Met Ser Ser Arg Tyr Asn Ala Phe Thr
                405                 410                 415
Arg Pro Ala Val Val Ser Val Arg Ala Gly Ser Ser Arg Leu Met Leu
            420                 425                 430
Arg Arg Glu Thr Leu Asp Asp Ile Leu Ser Leu Glu Ala
        435                 440                 445
```

(2) INFORMATION FOR SEQ ID NO: 6

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 20 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: other nucleic acid

(A) DESCRIPTION: /desc="Synthetic DNA"

(iv) ANTI-SENSE: NO

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:6:

CATCTAAGTA TGCATCTCGG 20

(2) INFORMATION FOR SEQ ID NO: 7

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 20 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: other nucleic acid

(A) DESCRIPTION: /desc="Synthetic DNA"

(iv) ANTI-SENSE: YES

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:7:

TGCCCCTCGA GCTAAATTAG 20

(2) INFORMATION FOR SEQ ID NO: 8

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 1034 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: double

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: Genomic DNA

(vi) ORIGINAL SOURCE:

(A) ORGANISM: Brevibacterium lactofermentum

(B) STRAIN: ATCC 13869

(ix) FEATURE:

(A) NAME/KEY: CDS

(B) LOCATION: 61..1020

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:8:

ATGCATCTCG GTAAGCTCGA CCAGGACAGT GCCACCACAA TTTTGGAGGA TTACAAGAAC 60

ATG ACC AAC ATC CGC GTA GCT ATC GTG GGC TAC GGA AAC CTG GGA CGC 108
Met Thr Asn Ile Arg Val Ala Ile Val Gly Tyr Gly Asn Leu Gly Arg
1 5 10 15

AGC GTC GAA AAG CTT ATT GCC AAG CAG CCC GAC ATG GAC CTT GTA GGA 156
Ser Val Glu Lys Leu Ile Ala Lys Gln Pro Asp Met Asp Leu Val Gly
20 25 30

ATC TTC TCG CGC CGG GCC ACC CTC GAC ACA AAG ACG CCA GTC TTT GAT 204
Ile Phe Ser Arg Arg Ala Thr Leu Asp Thr Lys Thr Pro Val Phe Asp
35 40 45

```
GTC GCC GAC GTG GAC AAG CAC GCC GAC GAC GTG GAC GTG CTG TTC CTG      252
Val Ala Asp Val Asp Lys His Ala Asp Asp Val Asp Val Leu Phe Leu
     50                  55                  60
TGC ATG GGC TCC GCC ACC GAC ATC CCT GAG CAG GCA CCA AAG TTC GCG      300
Cys Met Gly Ser Ala Thr Asp Ile Pro Glu Gln Ala Pro Lys Phe Ala
 65                  70                  75                  80
CAG TTC GCC TGC ACC GTA GAC ACC TAC GAC AAC CAC CGC GAC ATC CCA      348
Gln Phe Ala Cys Thr Val Asp Thr Tyr Asp Asn His Arg Asp Ile Pro
                 85                  90                  95
CGC CAC CGC CAG GTC ATG AAC GAA GCC GCC ACC GCA GCC GGC AAC GTT      396
Arg His Arg Gln Val Met Asn Glu Ala Ala Thr Ala Ala Gly Asn Val
            100                 105                 110
GCA CTG GTC TCT ACC GGC TGG GAT CCA GGA ATG TTC TCC ATC AAC CGC      444
Ala Leu Val Ser Thr Gly Trp Asp Pro Gly Met Phe Ser Ile Asn Arg
        115                 120                 125
GTC TAC GCA GCG GCA GTC TTA GCC GAG CAC CAG CAG CAC ACC TTC TGG      492
Val Tyr Ala Ala Ala Val Leu Ala Glu His Gln Gln His Thr Phe Trp
    130                 135                 140
GGC CCA GGT TTG TCA CAG GGC CAC TCC GAT GCT TTG CGA CGC ATC CCT      540
Gly Pro Gly Leu Ser Gln Gly His Ser Asp Ala Leu Arg Arg Ile Pro
145                 150                 155                 160
GGC GTT CAA AAG GCA GTC CAG TAC ACC CTC CCA TCC GAA GAC GCC CTG      588
Gly Val Gln Lys Ala Val Gln Tyr Thr Leu Pro Ser Glu Asp Ala Leu
                165                 170                 175
GAA AAG GCC CGC CGC GGC GAA GCC GGC GAC CTT ACC GGA AAG CAA ACC      636
Glu Lys Ala Arg Arg Gly Glu Ala Gly Asp Leu Thr Gly Lys Gln Thr
            180                 185                 190
CAC AAG CGC CAA TGC TTC GTG GTT GCC GAC GCG GCC GAT CAC GAG CGC      684
His Lys Arg Gln Cys Phe Val Val Ala Asp Ala Ala Asp His Glu Arg
        195                 200                 205
ATC GAA AAC GAC ATC CGC ACC ATG CCT GAT TAC TTC GTT GGC TAC GAA      732
Ile Glu Asn Asp Ile Arg Thr Met Pro Asp Tyr Phe Val Gly Tyr Glu
    210                 215                 220
GTC GAA GTC AAC TTC ATC GAC GAA GCA ACC TTC GAC TCC GAG CAC ACC      780
Val Glu Val Asn Phe Ile Asp Glu Ala Thr Phe Asp Ser Glu His Thr
225                 230                 235                 240
```

```
GGC ATG CCA CAC GGT GGC CAC GTG ATT ACC ACC GGC GAC ACC GGT GGC      828
Gly Met Pro His Gly Gly His Val Ile Thr Thr Gly Asp Thr Gly Gly
                245                 250                 255
TTC AAC CAC ACC GTG GAA TAC ATC CTC AAG CTG GAC CGA AAC CCA GAT      876
Phe Asn His Thr Val Glu Tyr Ile Leu Lys Leu Asp Arg Asn Pro Asp
            260                 265                 270
TTC ACC GCT TCC TCA CAG ATC GCT TTC GGT CGC GCA GCT CAC CGC ATG      924
Phe Thr Ala Ser Ser Gln Ile Ala Phe Gly Arg Ala Ala His Arg Met
        275                 280                 285
AAG CAG CAG GGC CAA AGC GGA GCT TTC ACC GTC CTC GAA GTT GCT CCA      972
Lys Gln Gln Gly Gln Ser Gly Ala Phe Thr Val Leu Glu Val Ala Pro
    290                 295                 300
TAC CTG CTC TCC CCA GAG AAC TTG GAC GAT CTG ATC GCA CGC GAC GTC     1020
Tyr Leu Leu Ser Pro Glu Asn Leu Asp Asp Leu Ile Ala Arg Asp Val
305                 310                 315                 320
TAATTTAGCT CGAG                                                    1034
```

(2) INFORMATION FOR SEQ ID NO: 9

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 320 amino acids

(B) TYPE: amino acid

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:9:

```
Met Thr Asn Ile Arg Val Ala Ile Val Gly Tyr Gly Asn Leu Gly Arg
 1               5                  10                  15
Ser Val Glu Lys Leu Ile Ala Lys Gln Pro Asp Met Asp Leu Val Gly
            20                  25                  30
Ile Phe Ser Arg Arg Ala Thr Leu Asp Thr Lys Thr Pro Val Phe Asp
        35                  40                  45
Val Ala Asp Val Asp Lys His Ala Asp Asp Val Asp Val Leu Phe Leu
    50                  55                  60
Cys Met Gly Ser Ala Thr Asp Ile Pro Glu Gln Ala Pro Lys Phe Ala
65                  70                  75                  80
Gln Phe Ala Cys Thr Val Asp Thr Tyr Asp Asn His Arg Asp Ile Pro
                85                  90                  95
```

```
Arg His Arg Gln Val Met Asn Glu Ala Ala Thr Ala Ala Gly Asn Val
            100                 105                 110
Ala Leu Val Ser Thr Gly Trp Asp Pro Gly Met Phe S r Ile Asn Arg
        115                 120                 125
Val Tyr Ala Ala Ala Val Leu Ala Glu His Gln Gln His Thr Phe Trp
    130                 135                 140
Gly Pro Gly Leu Ser Gln Gly His Ser Asp Ala Leu Arg Arg Ile Pro
145                 150                 155                 160
Gly Val Gln Lys Ala Val Gln Tyr Thr Leu Pro Ser Glu Asp Ala Leu
                165                 170                 175
Glu Lys Ala Arg Arg Gly Glu Ala Gly Asp Leu Thr Gly Lys Gln Thr
            180                 185                 190
His Lys Arg Gln Cys Phe Val Val Ala Asp Ala Ala Asp His Glu Arg
        195                 200                 205
Ile Glu Asn Asp Ile Arg Thr Met Pro Asp Tyr Phe Val Gly Tyr Glu
    210                 215                 220
Val Glu Val Asn Phe Ile Asp Glu Ala Thr Phe Asp Ser Glu His Thr
225                 230                 235                 240
Gly Met Pro His Gly Gly His Val Ile Thr Thr Gly Asp Thr Gly Gly
                245                 250                 255
Phe Asn His Thr Val Glu Tyr Ile Leu Lys Leu Asp Arg Asn Pro Asp
            260                 265                 270
Phe Thr Ala Ser Ser Gln Ile Ala Phe Gly Arg Ala Ala His Arg Met
        275                 280                 285
Lys Gln Gln Gly Gln Ser Gly Ala Phe Thr Val Leu Glu Val Ala Pro
    290                 295                 300
Tyr Leu Leu Ser Pro Glu Asn Leu Asp Asp Leu Ile Ala Arg Asp Val
305                 310                 315                 320
```

(2) INFORMATION FOR SEQ ID NO: 10

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 23 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: other nucleic acid

(A) DESCRIPTION: /desc="Synthetic DNA"

(iv) ANTI-SENSE: NO

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:10:

TCGCGAAGTA GCACCTGTCA CTT 23

(2) INFORMATION FOR SEQ ID NO: 11

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 21 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: other nucleic acid

(A) DESCRIPTION: /desc="Synthetic DNA"

(iv) ANTI-SENSE: YES

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:11:

ACGGAATTCA ATCTTACGGC C 21

(2) INFORMATION FOR SEQ ID NO: 12

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 1643 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: double

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: Genomic DNA

(vi) ORIGINAL SOURCE:

(A) ORGANISM: Brevibacterium lactofermentum

(B) STRAIN: ATCC 13869

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:12:

TCGCGAAGTA GCACCTGTCA CTTTTGTCTC AAATATTAAA TCGAATATCA ATATACGGTC 60

TGTTTATTGG AACGCATCCC AGTGGCTGAG ACGCATCCGC TAAAGCCCCA GGAACCCTGT 120

GCAGAAAGAA AACACTCCTC TGGCTAGGTA GACACAGTTT ATAAAGGTAG AGTTGAGCGG 180

GTAACTGTCA GCACGTAGAT CGAAAGGTGC ACAAAGGTGG CCCTGGTCGT ACAGAAATAT 240

GGCGGTTCCT CGCTTGAGAG TGCGGAACGC ATTAGAAACG TCGCTGAACG GATCGTTGCC 300

ACCAAGAAGG CTGGAAATGA TGTCGTGGTT GTCTGCTCCG CAATGGGAGA CACCACGGAT 360

GAACTTCTAG AACTTGCAGC GGCAGTGAAT CCCGTTCCGC CAGCTCGTGA AATGGATATG 420

CTCCTGACTG CTGGTGAGCG TATTTCTAAC GCTCTCGTCG CCATGGCTAT TGAGTCCCTT 480

GGCGCAGAAG CTCAATCTTT CACTGGCTCT CAGGCTGGTG TGCTCACCAC CGAGCGCCAC 540
GGAAACGCAC GCATTGTTGA CGTCACACCG GGTCGTGTGC GTGAAGCACT CGATGAGGGC 600
AAGATCTGCA TTGTTGCTGG TTTTCAGGGT GTTAATAAAG AAACCCGCGA TGTCACCACG 660
TTGGGTCGTG GTGGTTCTGA CACCACTGCA GTTGCGTTGG CAGCTGCTTT GAACGCTGAT 720
GTGTGTGAGA TTTACTCGGA CGTTGACGGT GTGTATACCG CTGACCCGCG CATCGTTCCT 780
AATGCACAGA AGCTGGAAAA GCTCAGCTTC GAAGAAATGC TGGAACTTGC TGCTGTTGGC 840
TCCAAGATTT TGGTGCTGCG CAGTGTTGAA TACGCTCGTG CATTCAATGT GCCACTTCGC 900
GTACGCTCGT CTTATAGTAA TGATCCCGGC ACTTTGATTG CCGGCTCTAT GGAGGATATT 960
CCTGTGGAAG AAGCAGTCCT TACCGGTGTC GCAACCGACA AGTCCGAAGC CAAAGTAACC 1020
GTTCTGGGTA TTTCCGATAA GCCAGGCGAG GCTGCCAAGG TTTTCCGTGC GTTGGCTGAT 1080
GCAGAAATCA ACATTGACAT GGTTCTGCAG AACGTCTCCT CTGTGGAAGA CGGCACCACC 1140
GACATCACGT TCACCTGCCC TCGCGCTGAC GGACGCCGTG CGATGGAGAT CTTGAAGAAG 1200
CTTCAGGTTC AGGGCAACTG GACCAATGTG CTTTACGACG ACCAGGTCGG CAAAGTCTCC 1260
CTCGTGGGTG CTGGCATGAA GTCTCACCCA GGTGTTACCG CAGAGTTCAT GGAAGCTCTG 1320
CGCGATGTCA ACGTGAACAT CGAATTGATT TCCACCTCTG AGATCCGCAT TTCCGTGCTG 1380
ATCCGTGAAG ATGATCTGGA TGCTGCTGCA CGTGCATTGC ATGAGCAGTT CCAGCTGGGC 1440
GGCGAAGACG AAGCCGTCGT TTATGCAGGC ACCGGACGCT AAAGTTTTAA AGGAGTAGTT 1500
TTACAATGAC CACCATCGCA GTTGTTGGTG CAACCGGCCA GGTCGGCCAG GTTATGCGCA 1560
CCCTTTTGGA AGAGCGCAAT TTCCCAGCTG ACACTGTTCG TTTCTTTGCT TCCCCGCGTT 1620
CCGCAGGCCG TAAGATTGAA TTC 1643

(2) INFORMATION FOR SEQ ID NO: 13

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 1643 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: double

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: Genomic DNA

(vi) ORIGINAL SOURCE:

(A) ORGANISM: Brevibacterium lactofermentum

(B) STRAIN: ATCC 13869

(ix) FEATURE:

(A) NAME/KEY: CDS

(B) LOCATION: 217..1482

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:13:

TCGCGAAGTA GCACCTGTCA CTTTTGTCTC AAATATTAAA TCGAATATCA ATATACGGTC 60

```
TGTTTATTGG AACGCATCCC AGTGGCTGAG ACGCATCCGC TAAAGCCCCA GGAACCCTGT    120
GCAGAAAGAA AACACTCCTC TGGCTAGGTA GACACAGTTT ATAAAGGTAG AGTTGAGCGG    180
GTAACTGTCA GCACGTAGAT CGAAAGGTGC ACAAAG GTG GCC CTG GTC GTA CAG      234
                                         Met Ala Leu Val Val Gln
                                          1               5
AAA TAT GGC GGT TCC TCG CTT GAG AGT GCG GAA CGC ATT AGA AAC GTC      282
Lys Tyr Gly Gly Ser Ser Leu Glu Ser Ala Glu Arg Ile Arg Asn Val
            10                  15                  20
GCT GAA CGG ATC GTT GCC ACC AAG AAG GCT GGA AAT GAT GTC GTG GTT      330
Ala Glu Arg Ile Val Ala Thr Lys Lys Ala Gly Asn Asp Val Val Val
        25                  30                  35
GTC TGC TCC GCA ATG GGA GAC ACC ACG GAT GAA CTT CTA GAA CTT GCA      378
Val Cys Ser Ala Met Gly Asp Thr Thr Asp Glu Leu Leu Glu Leu Ala
    40                  45                  50
GCG GCA GTG AAT CCC GTT CCG CCA GCT CGT GAA ATG GAT ATG CTC CTG      426
Ala Ala Val Asn Pro Val Pro Pro Ala Arg Glu Met Asp Met Leu Leu
55                  60                  65                  70
ACT GCT GGT GAG CGT ATT TCT AAC GCT CTC GTC GCC ATG GCT ATT GAG      474
Thr Ala Gly Glu Arg Ile Ser Asn Ala Leu Val Ala Met Ala Ile Glu
                75                  80                  85
TCC CTT GGC GCA GAA GCT CAA TCT TTC ACT GGC TCT CAG GCT GGT GTG      522
Ser Leu Gly Ala Glu Ala Gln Ser Phe Thr Gly Ser Gln Ala Gly Val
            90                  95                  100
CTC ACC ACC GAG CGC CAC GGA AAC GCA CGC ATT GTT GAC GTC ACA CCG      570
Leu Thr Thr Glu Arg His Gly Asn Ala Arg Ile Val Asp Val Thr Pro
        105                 110                 115
GGT CGT GTG CGT GAA GCA CTC GAT GAG GGC AAG ATC TGC ATT GTT GCT      618
Gly Arg Val Arg Glu Ala Leu Asp Glu Gly Lys Ile Cys Ile Val Ala
    120                 125                 130
GGT TTT CAG GGT GTT AAT AAA GAA ACC CGC GAT GTC ACC ACG TTG GGT      666
Gly Phe Gln Gly Val Asn Lys Glu Thr Arg Asp Val Thr Thr Leu Gly
135                 140                 145                 150
CGT GGT GGT TCT GAC ACC ACT GCA GTT GCG TTG GCA GCT GCT TTG AAC      714
Arg Gly Gly Ser Asp Thr Thr Ala Val Ala Leu Ala Ala Ala Leu Asn
                155                 160                 165
```

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GCT GAT GTG TGT GAG ATT TAC TCG GAC GTT GAC GGT GTG TAT ACC GCT      762
Ala Asp Val Cys Glu Ile Tyr Ser Asp Val Asp Gly Val Tyr Thr Ala
            170                 175                 180
GAC CCG CGC ATC GTT CCT AAT GCA CAG AAG CTG GAA AAG CTC AGC TTC      810
Asp Pro Arg Ile Val Pro Asn Ala Gln Lys Leu Glu Lys Leu Ser Phe
        185                 190                 195
GAA GAA ATG CTG GAA CTT GCT GCT GTT GGC TCC AAG ATT TTG GTG CTG      858
Glu Glu Met Leu Glu Leu Ala Ala Val Gly Ser Lys Ile Leu Val Leu
    200                 205                 210
CGC AGT GTT GAA TAC GCT CGT GCA TTC AAT GTG CCA CTT CGC GTA CGC      906
Arg Ser Val Glu Tyr Ala Arg Ala Phe Asn Val Pro Leu Arg Val Arg
215                 220                 225                 230
TCG TCT TAT AGT AAT GAT CCC GGC ACT TTG ATT GCC GGC TCT ATG GAG      954
Ser Ser Tyr Ser Asn Asp Pro Gly Thr Leu Ile Ala Gly Ser Met Glu
                235                 240                 245
GAT ATT CCT GTG GAA GAA GCA GTC CTT ACC GGT GTC GCA ACC GAC AAG     1002
Asp Ile Pro Val Glu Glu Ala Val Leu Thr Gly Val Ala Thr Asp Lys
            250                 255                 260
TCC GAA GCC AAA GTA ACC GTT CTG GGT ATT TCC GAT AAG CCA GGC GAG     1050
Ser Glu Ala Lys Val Thr Val Leu Gly Ile Ser Asp Lys Pro Gly Glu
        265                 270                 275
GCT GCC AAG GTT TTC CGT GCG TTG GCT GAT GCA GAA ATC AAC ATT GAC     1098
Ala Ala Lys Val Phe Arg Ala Leu Ala Asp Ala Glu Ile Asn Ile Asp
    280                 285                 290
ATG GTT CTG CAG AAC GTC TCC TCT GTG GAA GAC GGC ACC ACC GAC ATC     1146
Met Val Leu Gln Asn Val Ser Ser Val Glu Asp Gly Thr Thr Asp Ile
295                 300                 305                 310
ACG TTC ACC TGC CCT CGC GCT GAC GGA CGC CGT GCG ATG GAG ATC TTG     1194
Thr Phe Thr Cys Pro Arg Ala Asp Gly Arg Arg Ala Met Glu Ile Leu
                315                 320                 325
AAG AAG CTT CAG GTT CAG GGC AAC TGG ACC AAT GTG CTT TAC GAC GAC     1242
Lys Lys Leu Gln Val Gln Gly Asn Trp Thr Asn Val Leu Tyr Asp Asp
            330                 335                 340
CAG GTC GGC AAA GTC TCC CTC GTG GGT GCT GGC ATG AAG TCT CAC CCA     1290
Gln Val Gly Lys Val Ser Leu Val Gly Ala Gly Met Lys Ser His Pro
        345                 350                 355
```

```
GGT GTT ACC GCA GAG TTC ATG GAA GCT CTG CGC GAT GTC AAC GTG AAC      1338
Gly Val Thr Ala Glu Phe Met Glu Ala Leu Arg Asp Val Asn Val Asn
    360                 365                 370
ATC GAA TTG ATT TCC ACC TCT GAG ATC CGC ATT TCC GTG CTG ATC CGT      1386
Ile Glu Leu Ile Ser Thr Ser Glu Ile Arg Ile Ser Val Leu Ile Arg
375                 380                 385                 390
GAA GAT GAT CTG GAT GCT GCT GCA CGT GCA TTG CAT GAG CAG TTC CAG      1434
Glu Asp Asp Leu Asp Ala Ala Ala Arg Ala Leu His Glu Gln Phe Gln
                395                 400                 405
CTG GGC GGC GAA GAC GAA GCC GTC GTT TAT GCA GGC ACC GGA CGC TAA      1482
Leu Gly Gly Glu Asp Glu Ala Val Val Tyr Ala Gly Thr Gly Arg
            410                 415                 420
AGTTTTAAAG GAGTAGTTTT ACAATGACCA CCATCGCAGT TGTTGGTGCA ACCGGCCAGG    1542
TCGGCCAGGT TATGCGCACC CTTTTGGAAG AGCGCAATTT CCCAGCTGAC ACTGTTCGTT    1602
TCTTTGCTTC CCCGCGTTCC GCAGGCCGTA AGATTGAATT C                        1643
```

(2) INFORMATION FOR SEQ ID NO: 14

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 421 amino acids

(B) TYPE: amino acid

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:14:

```
Met Ala Leu Val Val Gln Lys Tyr Gly Gly Ser Ser Leu Glu Ser Ala
 1               5                  10                  15
Glu Arg Ile Arg Asn Val Ala Glu Arg Ile Val Ala Thr Lys Lys Ala
            20                  25                  30
Gly Asn Asp Val Val Val Val Cys Ser Ala Met Gly Asp Thr Thr Asp
        35                  40                  45
Glu Leu Leu Glu Leu Ala Ala Ala Val Asn Pro Val Pro Pro Ala Arg
    50                  55                  60
Glu Met Asp Met Leu Leu Thr Ala Gly Glu Arg Ile Ser Asn Ala Leu
65                  70                  75                  80
Val Ala Met Ala Ile Glu Ser Leu Gly Ala Glu Ala Gln Ser Phe Thr
                85                  90                  95
```

Gly Ser Gln Ala Gly Val Leu Thr Thr Glu Arg His Gly Asn Ala Arg
100 105 110
Ile Val Asp Val Thr Pro Gly Arg Val Arg Glu Ala Leu Asp Glu Gly
115 120 125
Lys Ile Cys Ile Val Ala Gly Phe Gln Gly Val Asn Lys Glu Thr Arg
130 135 140
Asp Val Thr Thr Leu Gly Arg Gly Gly Ser Asp Thr Thr Ala Val Ala
145 150 155 160
Leu Ala Ala Ala Leu Asn Ala Asp Val Cys Glu Ile Tyr Ser Asp Val
165 170 175
Asp Gly Val Tyr Thr Ala Asp Pro Arg Ile Val Pro Asn Ala Gln Lys
180 185 190
Leu Glu Lys Leu Ser Phe Glu Glu Met Leu Glu Leu Ala Ala Val Gly
195 200 205
Ser Lys Ile Leu Val Leu Arg Ser Val Glu Tyr Ala Arg Ala Phe Asn
210 215 220
Val Pro Leu Arg Val Arg Ser Ser Tyr Ser Asn Asp Pro Gly Thr Leu
225 230 235 240
Ile Ala Gly Ser Met Glu Asp Ile Pro Val Glu Glu Ala Val Leu Thr
245 250 255
Gly Val Ala Thr Asp Lys Ser Glu Ala Lys Val Thr Val Leu Gly Ile
260 265 270
Ser Asp Lys Pro Gly Glu Ala Ala Lys Val Phe Arg Ala Leu Ala Asp
275 280 285
Ala Glu Ile Asn Ile Asp Met Val Leu Gln Asn Val Ser Ser Val Glu
290 295 300
Asp Gly Thr Thr Asp Ile Thr Phe Thr Cys Pro Arg Ala Asp Gly Arg
305 310 315 320
Arg Ala Met Glu Ile Leu Lys Lys Leu Gln Val Gln Gly Asn Trp Thr
325 330 335
Asn Val Leu Tyr Asp Asp Gln Val Gly Lys Val Ser Leu Val Gly Ala
340 345 350
Gly Met Lys Ser His Pro Gly Val Thr Ala Glu Phe Met Glu Ala Leu
355 360 365
Arg Asp Val Asn Val Asn Ile Glu Leu Ile Ser Thr Ser Glu Ile Arg
370 375 380

Ile Ser Val Leu Ile Arg Glu Asp Asp Leu Asp Ala Ala Ala Arg Ala
385 390 395 400
Leu His Glu Gln Phe Gln Leu Gly Gly Glu Asp Glu Ala Val Val Tyr
405 410 415
Ala Gly Thr Gly Arg
420

(2) INFORMATION FOR SEQ ID NO: 15

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 1643 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: double

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: Genomic DNA

(vi) ORIGINAL SOURCE:

(A) ORGANISM: Brevibacterium lactofermentum

(B) STRAIN: ATCC 13869

(ix) FEATURE:

(A) NAME/KEY: CDS

(B) LOCATION: 964..1482

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:15:

TCGCGAAGTA GCACCTGTCA CTTTTGTCTC AAATATTAAA TCGAATATCA ATATACGGTC 60
TGTTTATTGG AACGCATCCC AGTGGCTGAG ACGCATCCGC TAAAGCCCCA GGAACCCTGT 120
GCAGAAAGAA AACACTCCTC TGGCTAGGTA GACACAGTTT ATAAAGGTAG AGTTGAGCGG 180
GTAACTGTCA GCACGTAGAT CGAAAGGTGC ACAAAGGTGG CCCTGGTCGT ACAGAAATAT 240
GGCGGTTCCT CGCTTGAGAG TGCGGAACGC ATTAGAAACG TCGCTGAACG GATCGTTGCC 300
ACCAAGAAGG CTGGAAATGA TGTCGTGGTT GTCTGCTCCG CAATGGGAGA CACCACGGAT 360
GAACTTCTAG AACTTGCAGC GGCAGTGAAT CCCGTTCCGC CAGCTCGTGA AATGGATATG 420
CTCCTGACTG CTGGTGAGCG TATTTCTAAC GCTCTCGTCG CCATGGCTAT TGAGTCCCTT 480
GGCGCAGAAG CTCAATCTTT CACTGGCTCT CAGGCTGGTG TGCTCACCAC CGAGCGCCAC 540
GGAAACGCAC GCATTGTTGA CGTCACACCG GGTCGTGTGC GTGAAGCACT CGATGAGGGC 600
AAGATCTGCA TTGTTGCTGG TTTTCAGGGT GTTAATAAAG AAACCCGCGA TGTCACCACG 660
TTGGGTCGTG GTGGTTCTGA CACCACTGCA GTTGCGTTGG CAGCTGCTTT GAACGCTGAT 720
GTGTGTGAGA TTTACTCGGA CGTTGACGGT GTGTATACCG CTGACCCGCG CATCGTTCCT 780
AATGCACAGA AGCTGGAAAA GCTCAGCTTC GAAGAAATGC TGGAACTTGC TGCTGTTGGC 840
TCCAAGATTT TGGTGCTGCG CAGTGTTGAA TACGCTCGTG CATTCAATGT GCCACTTCGC 900

```
GTACGCTCGT CTTATAGTAA TGATCCCGGC ACTTTGATTG CCGGCTCTAT GGAGGATATT    960
CCT  GTG GAA GAA GCA GTC CTT ACC GGT GTC GCA ACC GAC AAG TCC GAA    1008
     Met Glu Glu Ala Val Leu Thr Gly Val Ala Thr Asp Lys Ser Glu
       1               5                  10                  15
GCC AAA GTA ACC GTT CTG GGT ATT TCC GAT AAG CCA GGC GAG GCT GCC    1056
Ala Lys Val Thr Val Leu Gly Ile Ser Asp Lys Pro Gly Glu Ala Ala
                   20                  25                  30
AAG GTT TTC CGT GCG TTG GCT GAT GCA GAA ATC AAC ATT GAC ATG GTT    1104
Lys Val Phe Arg Ala Leu Ala Asp Ala Glu Ile Asn Ile Asp Met Val
               35                  40                  45
CTG CAG AAC GTC TCC TCT GTG GAA GAC GGC ACC ACC GAC ATC ACG TTC    1152
Leu Gln Asn Val Ser Ser Val Glu Asp Gly Thr Thr Asp Ile Thr Phe
           50                  55                  60
ACC TGC CCT CGC GCT GAC GGA CGC CGT GCG ATG GAG ATC TTG AAG AAG    1200
Thr Cys Pro Arg Ala Asp Gly Arg Arg Ala Met Glu Ile Leu Lys Lys
       65                  70                  75
CTT CAG GTT CAG GGC AAC TGG ACC AAT GTG CTT TAC GAC GAC CAG GTC    1248
Leu Gln Val Gln Gly Asn Trp Thr Asn Val Leu Tyr Asp Asp Gln Val
80                  85                  90                  95
GGC AAA GTC TCC CTC GTG GGT GCT GGC ATG AAG TCT CAC CCA GGT GTT    1296
Gly Lys Val Ser Leu Val Gly Ala Gly Met Lys Ser His Pro Gly Val
                100                 105                 110
ACC GCA GAG TTC ATG GAA GCT CTG CGC GAT GTC AAC GTG AAC ATC GAA    1344
Thr Ala Glu Phe Met Glu Ala Leu Arg Asp Val Asn Val Asn Ile Glu
            115                 120                 125
TTG ATT TCC ACC TCT GAG ATC CGC ATT TCC GTG CTG ATC CGT GAA GAT    1392
Leu Ile Ser Thr Ser Glu Ile Arg Ile Ser Val Leu Ile Arg Glu Asp
        130                 135                 140
GAT CTG GAT GCT GCT GCA CGT GCA TTG CAT GAG CAG TTC CAG CTG GGC    1440
Asp Leu Asp Ala Ala Ala Arg Ala Leu His Glu Gln Phe Gln Leu Gly
    145                 150                 155
GGC GAA GAC GAA GCC GTC GTT TAT GCA GGC ACC GGA CGC TAAAGTTTTAA    1490
Gly Glu Asp Glu Ala Val Val Tyr Ala Gly Thr Gly Arg
160                 165                 170
AGGAGTAGTT TTACAATGAC CACCATCGCA GTTGTTGGTG CAACCGGCCA GGTCGGCCAG  1550
GTTATGCGCA CCCTTTTGGA AGAGCGCAAT TTCCCAGCTG ACACTGTTCG TTTCTTTGCT  1610
```

TCCCCGCGTT CCGCAGGCCG TAAGATTGAA TTC 1643

(2) INFORMATION FOR SEQ ID NO: 16

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 172 amino acids

(B) TYPE: amino acid

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:16:

Met Glu Glu Ala Val Leu Thr Gly Val Ala Thr Asp Lys Ser Glu Ala
1 5 10 15
Lys Val Thr Val Leu Gly Ile Ser Asp Lys Pro Gly Glu Ala Ala Lys
20 25 30
Val Phe Arg Ala Leu Ala Asp Ala Glu Ile Asn Ile Asp Met Val Leu
35 40 45
Gln Asn Val Ser Ser Val Glu Asp Gly Thr Thr Asp Ile Thr Phe Thr
50 55 60
Cys Pro Arg Ala Asp Gly Arg Arg Ala Met Glu Ile Leu Lys Lys Leu
65 70 75 80
Gln Val Gln Gly Asn Trp Thr Asn Val Leu Tyr Asp Asp Gln Val Gly
85 90 95
Lys Val Ser Leu Val Gly Ala Gly Met Lys Ser His Pro Gly Val Thr
100 105 110
Ala Glu Phe Met Glu Ala Leu Arg Asp Val Asn Val Asn Ile Glu Leu
115 120 125
Ile Ser Thr Ser Glu Ile Arg Ile Ser Val Leu Ile Arg Glu Asp Asp
130 135 140
Leu Asp Ala Ala Ala Arg Ala Leu His Glu Gln Phe Gln Leu Gly Gly
145 150 155 160
Glu Asp Glu Ala Val Val Tyr Ala Gly Thr Gly Arg
165 170

(2) INFORMATION FOR SEQ ID NO: 17

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 23 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single
(D) TOPOLOGY: linear
(ii) MOLECULE TYPE: other nucleic acid
(A) DESCRIPTION: /desc="Synthetic DNA"
(iv) ANTI-SENSE: NO
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:17:

GTCGACGGAT CGCAAATGGC AAC 23

(2) INFORMATION FOR SEQ ID NO: 18
(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 23 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear
(ii) MOLECULE TYPE: other nucleic acid
(A) DESCRIPTION: /desc="Synthetic DNA"
(iv) ANTI-SENSE: YES
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:18:

GGATCCTTGA GCACCTTGCG CAG 23

(2) INFORMATION FOR SEQ ID NO: 19
(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 1411 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: double
(D) TOPOLOGY: linear
(ii) MOLECULE TYPE: Genomic DNA
(vi) ORIGINAL SOURCE:
(A) ORGANISM: Brevibacterium lactofermentum
(B) STRAIN: ATCC 13869
(ix) FEATURE:
(A) NAME/KEY: CDS
(B) LOCATION: 311..1213
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:19:

CTCTCGATAT CGAGAGAGAA GCAGCGCCAC GGTTTTTCGG TGATTTTGAG ATTGAAACTT 60
TGGCAGACGG ATCGCAAATG GCAACAAGCC CGTATGTCAT GGACTTTTAA CGCAAAGCTC 120

```
ACACCCACGA GCTAAAAATT CATATAGTTA AGACAACATT TTTGGCTGTA AAAGACAGCC   180
GTAAAAACCT CTTGCTCATG TCAATTGTTC TTATCGGAAT GTGGCTTGGG CGATTGTTAT   240
GCAAAAGTTG TTAGGTTTTT TGCGGGGTTG TTTAACCCCC AAATGAGGGA AGAAGGTAAC   300
CTTGAACTCT ATG AGC ACA GGT TTA ACA GCT AAG ACC GGA GTA GAG CAC       349
           Met Ser Thr Gly Leu Thr Ala Lys Thr Gly Val Glu His
            1               5                  10
TTC GGC ACC GTT GGA GTA GCA ATG GTT ACT CCA TTC ACG GAA TCC GGA      397
Phe Gly Thr Val Gly Val Ala Met Val Thr Pro Phe Thr Glu Ser Gly
     15                  20                  25
GAC ATC GAT ATC GCT GCT GGC CGC GAA GTC GCG GCT TAT TTG GTT GAT      445
Asp Ile Asp Ile Ala Ala Gly Arg Glu Val Ala Ala Tyr Leu Val Asp
 30                  35                  40                  45
AAG GGC TTG GAT TCT TTG GTT CTC GCG GGC ACC ACT GGT GAA TCC CCA      493
Lys Gly Leu Asp Ser Leu Val Leu Ala Gly Thr Thr Gly Glu Ser Pro
                 50                  55                  60
ACG ACA ACC GCC GCT GAA AAA CTA GAA CTG CTC AAG GCC GTT CGT GAG      541
Thr Thr Thr Ala Ala Glu Lys Leu Glu Leu Leu Lys Ala Val Arg Glu
            65                  70                  75
GAA GTT GGG GAT CGG GCG AAC GTC ATC GCC GGT GTC GGA ACC AAC AAC      589
Glu Val Gly Asp Arg Ala Asn Val Ile Ala Gly Val Gly Thr Asn Asn
         80                  85                  90
ACG CGG ACA TCT GTG GAA CTT GCG GAA GCT GCT GCT TCT GCT GGC GCA      637
Thr Arg Thr Ser Val Glu Leu Ala Glu Ala Ala Ala Ser Ala Gly Ala
     95                  100                 105
GAC GGC CTT TTA GTT GTA ACT CCT TAT TAC TCC AAG CCG AGC CAA GAG      685
Asp Gly Leu Leu Val Val Thr Pro Tyr Tyr Ser Lys Pro Ser Gln Glu
110                 115                 120                 125
GGA TTG CTG GCG CAC TTC GGT GCA ATT GCT GCA GCA ACA GAG GTT CCA      733
Gly Leu Leu Ala His Phe Gly Ala Ile Ala Ala Ala Thr Glu Val Pro
                130                 135                 140
ATT TGT CTC TAT GAC ATT CCT GGT CGG TCA GGT ATT CCA ATT GAG TCT      781
Ile Cys Leu Tyr Asp Ile Pro Gly Arg Ser Gly Ile Pro Ile Glu Ser
            145                 150                 155
GAT ACC ATG AGA CGC CTG AGT GAA TTA CCT ACG ATT TTG GCG GTC AAG      829
Asp Thr Met Arg Arg Leu Ser Glu Leu Pro Thr Ile Leu Ala Val Lys
         160                 165                 170
```

```
GAC GCC AAG GGT GAC CTC GTT GCA GCC ACG TCA TTG ATC AAA GAA ACG      877
Asp Ala Lys Gly Asp Leu Val Ala Ala Thr Ser Leu Ile Lys Glu Thr
    175                 180                 185
GGA CTT GCC TGG TAT TCA GGC GAT GAC CCA CTA AAC CTT GTT TGG CTT      925
Gly Leu Ala Trp Tyr Ser Gly Asp Asp Pro Leu Asn Leu Val Trp Leu
190                 195                 200                 205
GCT TTG GGC GGA TCA GGT TTC ATT TCC GTA ATT GGA CAT GCA GCC CCC      973
Ala Leu Gly Gly Ser Gly Phe Ile Ser Val Ile Gly His Ala Ala Pro
                210                 215                 220
ACA GCA TTA CGT GAG TTG TAC ACA AGC TTC GAG GAA GGC GAC CTC GTC     1021
Thr Ala Leu Arg Glu Leu Tyr Thr Ser Phe Glu Glu Gly Asp Leu Val
            225                 230                 235
CGT GCG CGG GAA ATC AAC GCC AAA CTA TCA CCG CTG GTA GCT GCC CAA     1069
Arg Ala Arg Glu Ile Asn Ala Lys Leu Ser Pro Leu Val Ala Ala Gln
        240                 245                 250
GGT CGC TTG GGT GGA GTC AGC TTG GCA AAA GCT GCT CTG CGT CTG CAG     1117
Gly Arg Leu Gly Gly Val Ser Leu Ala Lys Ala Ala Leu Arg Leu Gln
    255                 260                 265
GGC ATC AAC GTA GGA GAT CCT CGA CTT CCA ATT ATG GCT CCA AAT GAG     1165
Gly Ile Asn Val Gly Asp Pro Arg Leu Pro Ile Met Ala Pro Asn Glu
270                 275                 280                 285
CAG GAA CTT GAG GCT CTC CGA GAA GAC ATG AAA AAA GCT GGA GTT CTA     1213
Gln Glu Leu Glu Ala Leu Arg Glu Asp Met Lys Lys Ala Gly Val Leu
                290                 295                 300
TAAATATGAA TGATTCCCGA AATCGCGGCC GGAAGGTTAC CCGCAAGGCG GCCCACCAGA   1273
AGCTGGTCAG GAAAACCATC TGGATACCCC TGTCTTTCAG GCACCAGATG CTTCCTCTAA   1333
CCAGAGCGCT GTAAAAGCTG AGACCGCCGG AAACGACAAT CGGGATGCTG CGCAAGGTGC   1393
TCAAGGATCC CAACATTC                                                 1411
```

(2) INFORMATION FOR SEQ ID NO: 20

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 301 amino acids

(B) TYPE: amino acid

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:20:

```
Met Ser Thr Gly Leu Thr Ala Lys Thr Gly Val Glu His Phe Gly Thr
 1               5                  10                  15
Val Gly Val Ala Met Val Thr Pro Phe Thr Glu Ser Gly Asp Ile Asp
            20                  25                  30
Ile Ala Ala Gly Arg Glu Val Ala Ala Tyr Leu Val Asp Lys Gly Leu
        35                  40                  45
Asp Ser Leu Val Leu Ala Gly Thr Thr Gly Glu Ser Pro Thr Thr Thr
    50                  55                  60
Ala Ala Glu Lys Leu Glu Leu Leu Lys Ala Val Arg Glu Glu Val Gly
65                  70                  75                  80
Asp Arg Ala Asn Val Ile Ala Gly Val Gly Thr Asn Asn Thr Arg Thr
                85                  90                  95
Ser Val Glu Leu Ala Glu Ala Ala Ala Ser Ala Gly Ala Asp Gly Leu
            100                 105                 110
Leu Val Val Thr Pro Tyr Tyr Ser Lys Pro Ser Gln Glu Gly Leu Leu
        115                 120                 125
Ala His Phe Gly Ala Ile Ala Ala Ala Thr Glu Val Pro Ile Cys Leu
    130                 135                 140
Tyr Asp Ile Pro Gly Arg Ser Gly Ile Pro Ile Glu Ser Asp Thr Met
145                 150                 155                 160
Arg Arg Leu Ser Glu Leu Pro Thr Ile Leu Ala Val Lys Asp Ala Lys
                165                 170                 175
Gly Asp Leu Val Ala Ala Thr Ser Leu Ile Lys Glu Thr Gly Leu Ala
            180                 185                 190
Trp Tyr Ser Gly Asp Asp Pro Leu Asn Leu Val Trp Leu Ala Leu Gly
        195                 200                 205
Gly Ser Gly Phe Ile Ser Val Ile Gly His Ala Ala Pro Thr Ala Leu
    210                 215                 220
Arg Glu Leu Tyr Thr Ser Phe Glu Glu Gly Asp Leu Val Arg Ala Arg
225                 230                 235                 240
Glu Ile Asn Ala Lys Leu Ser Pro Leu Val Ala Ala Gln Gly Arg Leu
                245                 250                 255
Gly Gly Val Ser Leu Ala Lys Ala Ala Leu Arg Leu Gln Gly Ile Asn
            260                 265                 270
Val Gly Asp Pro Arg Leu Pro Ile Met Ala Pro Asn Glu Gln Glu Leu
        275                 280                 285
```

Glu Ala Leu Arg Glu Asp Met Lys Lys Ala Gly Val Leu
290 295 300

(2) INFORMATION FOR SEQ ID NO: 21
(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 23 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear
(ii) MOLECULE TYPE: other nucleic acid
(A) DESCRIPTION: /desc="Synthetic DNA"
(iv) ANTI-SENSE: NO
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:21:
GGATCCCCAA TCGATACCTG GAA 23

(2) INFORMATION FOR SEQ ID NO: 22
(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 23 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear
(ii) MOLECULE TYPE: other nucleic acid
(A) DESCRIPTION: /desc="Synthetic DNA"
(iv) ANTI-SENSE: YES
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:22:
CGGTTCATCG CCAAGTTTTT CTT 23

(2) INFORMATION FOR SEQ ID NO: 23
(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 2001 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: double
(D) TOPOLOGY: linear
(ii) MOLECULE TYPE: Genomic DNA
(vi) ORIGINAL SOURCE:
(A) ORGANISM: Brevibacterium lactofermentum

(B) STRAIN: ATCC 13869

(ix) FEATURE:

(A) NAME/KEY: CDS

(B) LOCATION: 730..1473

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:23:

```
GGATCCCCAA TCGATACCTG GAACGACAAC CTGATCAGGA TATCCAATGC CTTGAATATT      60
GACGTTGAGG AAGGAATCAC CAGCCATCTC AACTGGAAGA CCTGACGCCT GCTGAATTGG     120
ATCAGTGGCC CAATCGACCC ACCAACCAGG TTGGCTATTA CCGGCGATAT CAAAAACAAC     180
TCGCGTGAAC GTTTCGTGCT CGGCAACGCG GATGCCAGCG ATCGACATAT CGGAGTCACC     240
AACTTGAGCC TGCTGCTTCT GATCCATCGA CGGGGAACCC AACGGCGGCA AAGCAGTGGG     300
GGAAGGGGAG TTGGTGGACT CTGAATCAGT GGGCTCTGAA GTGGTAGGCG ACGGGGCAGC     360
ATCTGAAGGC GTGCGAGTTG TGGTGACCGG GTTAGCGGTT TCAGTTTCTG TCACAACTGG     420
AGCAGGACTA GCAGAGGTTG TAGGCGTTGA GCCGCTTCCA TCACAAGCAC TTAAAAGTAA     480
AGAGGCGGAA ACCACAAGCG CCAAGGAACT ACCTGCGGAA CGGGCGGTGA AGGGCAACTT     540
AAGTCTCATA TTTCAAACAT AGTTCCACCT GTGTGATTAA TCTCCAGAAC GGAACAAACT     600
GATGAACAAT CGTTAACAAC ACAGACCAAA ACGGTCAGTT AGGTATGGAT ATCAGCACCT     660
TCTGAATGGG TACGTCTAGA CTGGTGGGCG TTTGAAAAAC TCTTCGCCCC ACGAAAATGA     720
AGGAGCATA ATG GGA ATC AAG GTT GGC GTT CTC GGA GCC AAA GGC CGT           768
          Met Gly Ile Lys Val Gly Val Leu Gly Ala Lys Gly Arg
            1               5                  10
GTT GGT CAA ACT ATT GTG GCA GCA GTC AAT GAG TCC GAC GAT CTG GAG        816
Val Gly Gln Thr Ile Val Ala Ala Val Asn Glu Ser Asp Asp Leu Glu
     15                  20                  25
CTT GTT GCA GAG ATC GGC GTC GAC GAT GAT TTG AGC CTT CTG GTA GAC        864
Leu Val Ala Glu Ile Gly Val Asp Asp Asp Leu Ser Leu Leu Val Asp
 30                  35                  40                  45
AAC GGC GCT GAA GTT GTC GTT GAC TTC ACC ACT CCT AAC GCT GTG ATG        912
Asn Gly Ala Glu Val Val Val Asp Phe Thr Thr Pro Asn Ala Val Met
                 50                  55                  60
GGC AAC CTG GAG TTC TGC ATC AAC AAC GGC ATT TCT GCG GTT GTT GGA        960
Gly Asn Leu Glu Phe Cys Ile Asn Asn Gly Ile Ser Ala Val Val Gly
             65                  70                  75
ACC ACG GGC TTC GAT GAT GCT CGT TTG GAG CAG GTT CGC GCC TGG CTT       1008
Thr Thr Gly Phe Asp Asp Ala Arg Leu Glu Gln Val Arg Ala Trp Leu
         80                  85                  90
```

```
GAA GGA AAA GAC AAT GTC GGT GTT CTG ATC GCA CCT AAC TTT GCT ATC      1056
Glu Gly Lys Asp Asn Val Gly Val Leu Ile Ala Pro Asn Phe Ala Ile
     95                  100                 105
TCT GCG GTG TTG ACC ATG GTC TTT TCC AAG CAG GCT GCC CGC TTC TTC      1104
Ser Ala Val Leu Thr Met Val Phe Ser Lys Gln Ala Ala Arg Phe Phe
110                 115                 120                 125
GAA TCA GCT GAA GTT ATT GAG CTG CAC CAC CCC AAC AAG CTG GAT GCA      1152
Glu Ser Ala Glu Val Ile Glu Leu His His Pro Asn Lys Leu Asp Ala
                130                 135                 140
CCT TCA GGC ACC GCG ATC CAC ACT GCT CAG GGC ATT GCT GCG GCA CGC      1200
Pro Ser Gly Thr Ala Ile His Thr Ala Gln Gly Ile Ala Ala Ala Arg
            145                 150                 155
AAA GAA GCA GGC ATG GAC GCA CAG CCA GAT GCG ACC GAG CAG GCA CTT      1248
Lys Glu Ala Gly Met Asp Ala Gln Pro Asp Ala Thr Glu Gln Ala Leu
        160                 165                 170
GAG GGT TCC CGT GGC GCA AGC GTA GAT GGA ATC CCA GTT CAC GCA GTC      1296
Glu Gly Ser Arg Gly Ala Ser Val Asp Gly Ile Pro Val His Ala Val
    175                 180                 185
CGC ATG TCC GGC ATG GTT GCT CAC GAG CAA GTT ATC TTT GGC ACC CAG      1344
Arg Met Ser Gly Met Val Ala His Glu Gln Val Ile Phe Gly Thr Gln
190                 195                 200                 205
GGT CAG ACC TTG ACC ATC AAG CAG GAC TCC TAT GAT CGC AAC TCA TTT      1392
Gly Gln Thr Leu Thr Ile Lys Gln Asp Ser Tyr Asp Arg Asn Ser Phe
                210                 215                 220
GCA CCA GGT GTC TTG GTG GGT GTG CGC AAC ATT GCA CAG CAC CCA GGC      1440
Ala Pro Gly Val Leu Val Gly Val Arg Asn Ile Ala Gln His Pro Gly
            225                 230                 235
CTA GTC GTA GGA CTT GAG CAT TAC CTA GGC CTG TAAAGGCTCA TTTCAGCAGC    1493
Leu Val Val Gly Leu Glu His Tyr Leu Gly Leu
        240                 245
GGGTGGAATT TTTTAAAAGG AGCGTTTAAA GGCTGTGGCC GAACAAGTTA AATTGAGCGT    1553
GGAGTTGATA GCGTGCAGTT CTTTTACTCC ACCCGCTGAT GTTGAGTGGT CAACTGATGT    1613
TGAGGGCGCG GAAGCACTCG TCGAGTTTGC GGGTCGTGCC TGCTACGAAA CTTTTGATAA    1673
GCCGAACCCT CGAACTGCTT CCAATGCTGC GTATCTGCGC CACATCATGG AAGTGGGGCA    1733
CACTGCTTTG CTTGAGCATG CCAATGCCAC GATGTATATC CGAGGCATTT CTCGGTCCGC    1793
GACCCATGAA TTGGTCCGAC ACCGCCATTT TTCCTTCTCT CAACTGTCTC AGCGTTTCGT    1853
```

GCACAGCGGA GAATCGGAAG TAGTGGTGCC CACTCTCATC GATGAAGATC CGCAGTTGCG 1913
TGAACTTTTC ATGCACGCCA TGGATGAGTC TCGGTTCGCT TTCAATGAGC TGCTTAATGC 1973
GCTGGAAGAA AAACTTGGCG ATGAACCG 2001

(2) INFORMATION FOR SEQ ID NO: 24

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 248 amino acids

(B) TYPE: amino acid

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:24:

```
Met Gly Ile Lys Val Gly Val Leu Gly Ala Lys Gly Arg Val Gly Gln
 1               5                  10                  15
Thr Ile Val Ala Ala Val Asn Glu Ser Asp Asp Leu Glu Leu Val Ala
            20                  25                  30
Glu Ile Gly Val Asp Asp Asp Leu Ser Leu Leu Val Asp Asn Gly Ala
        35                  40                  45
Glu Val Val Val Asp Phe Thr Thr Pro Asn Ala Val Met Gly Asn Leu
    50                  55                  60
Glu Phe Cys Ile Asn Asn Gly Ile Ser Ala Val Val Gly Thr Thr Gly
65                  70                  75                  80
Phe Asp Asp Ala Arg Leu Glu Gln Val Arg Ala Trp Leu Glu Gly Lys
                85                  90                  95
Asp Asn Val Gly Val Leu Ile Ala Pro Asn Phe Ala Ile Ser Ala Val
            100                 105                 110
Leu Thr Met Val Phe Ser Lys Gln Ala Ala Arg Phe Phe Glu Ser Ala
        115                 120                 125
Glu Val Ile Glu Leu His His Pro Asn Lys Leu Asp Ala Pro Ser Gly
    130                 135                 140
Thr Ala Ile His Thr Ala Gln Gly Ile Ala Ala Ala Arg Lys Glu Ala
145                 150                 155                 160
Gly Met Asp Ala Gln Pro Asp Ala Thr Glu Gln Ala Leu Glu Gly Ser
                165                 170                 175
Arg Gly Ala Ser Val Asp Gly Ile Pro Val His Ala Val Arg Met Ser
            180                 185                 190
Gly Met Val Ala His Glu Gln Val Ile Phe Gly Thr Gln Gly Gln Thr
```

```
                195                 200                 205
Leu Thr Ile Lys Gln Asp Ser Tyr Asp Arg Asn Ser Phe Ala Pro Gly
    210                 215                 220
Val Leu Val Gly Val Arg Asn Ile Ala Gln His Pro Gly Leu Val Val
225                 230                 235                 240
Gly Leu Glu His Tyr Leu Gly Leu
                245
```

Claims

1. A recombinant DNA autonomously replicable in cells of coryneform bacteria, comprising a DNA sequence coding for a diaminopicolinate decarboxylase, and a DNA sequence coding for a diaminopicolinate dehydrogenase.

2. The recombinant DNA according to claim 2, wherein said DNA sequence coding for the diaminopicolinate decarboxylase codes for an amino acid sequence shown in SEQ ID NO: 5 in Sequence Listing, or an amino acid sequence substantially the same as the amino acid sequence shown in SEQ ID NO: 5.

3. The recombinant DNA according to claim 1, wherein said DNA sequence coding for the diaminopicolinate dehydrogenase codes for an amino acid sequence depicted in SEQ ID NO: 9 in Sequence Listing, or an amino acid sequence substantially the same as the amino acid sequence depicted in SEQ ID NO: 9.

4. A coryneform bacterium in which said DNA sequence coding for a diaminopicolinate decarboxylase, and said DNA sequence coding for a diaminopicolinate dehydrogenase are enhanced.

5. The coryneform bacterium according to claim 4, which is transformed by introduction of the recombinant DNA as defined in claim 1.

6. A method for producing L-lysine comprising the steps of cultivating said coryneform bacterium as defined in claim 4 in a medium, to allow L-lysine to be produced and accumulated in a culture, and collecting L-lysine from the culture.

7. A method for producing L-lysine comprising the steps of cultivating said coryneform bacterium as defined in claim 5 in a medium, to allow L-lysine to be produced and accumulated in a culture, and collecting L-lysine from the culture.

FIG. 1

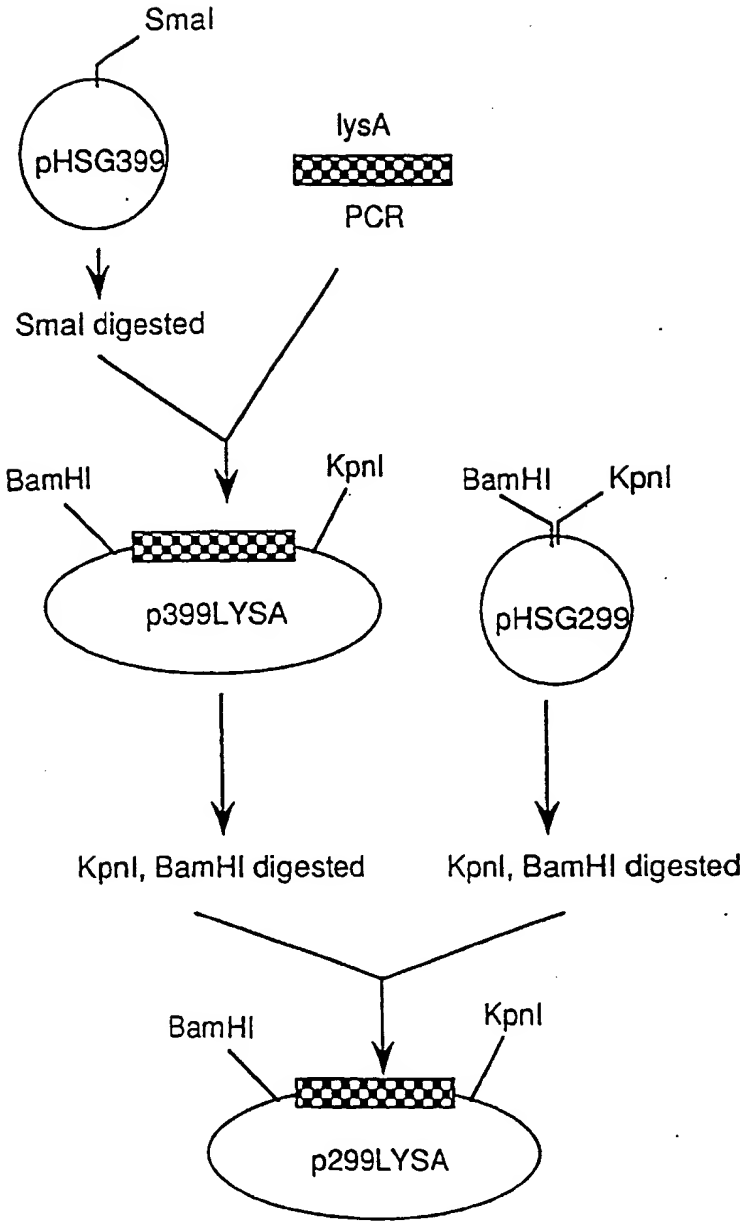

SmaI
pHSG399
lysA
PCR
SmaI digested
BamHI
KpnI
p399LYSA
BamHI
KpnI
pHSG299
KpnI, BamHI digested
KpnI, BamHI digested
BamHI
KpnI
p299LYSA

FIG. 2

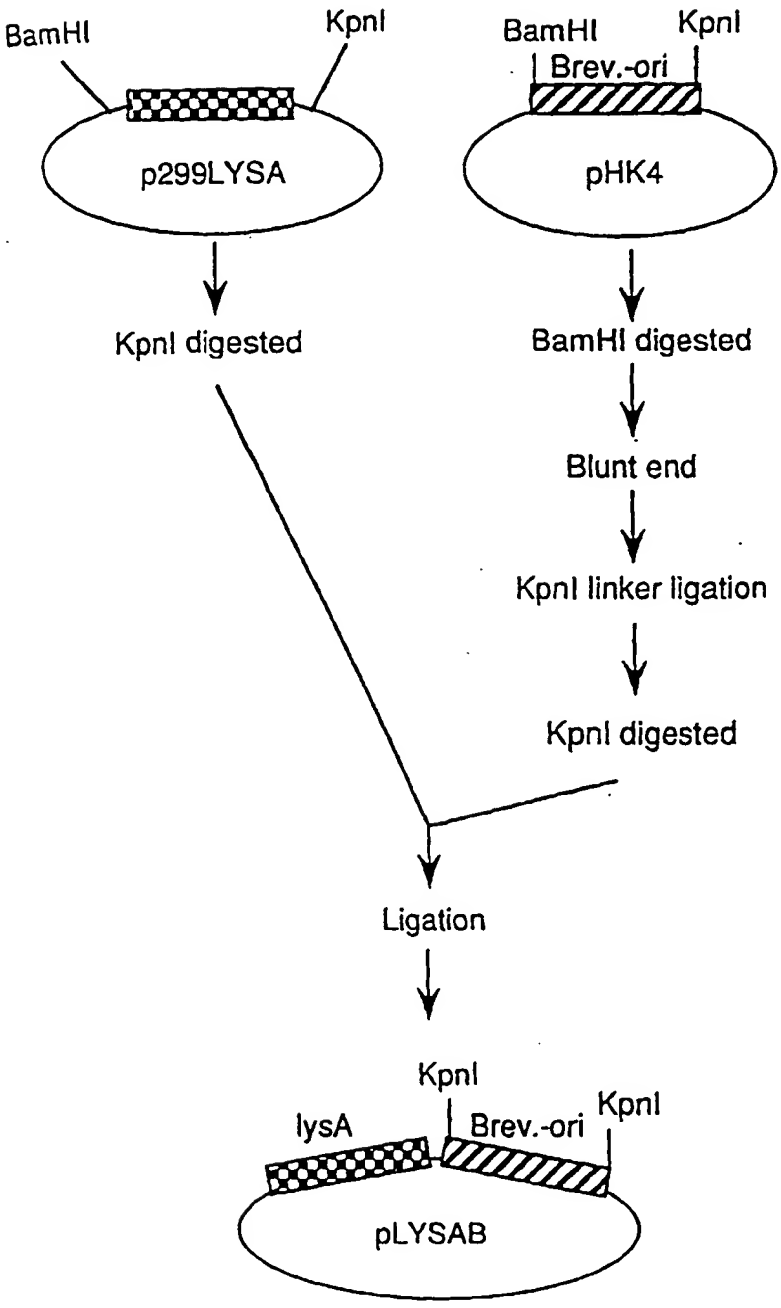

BamHI
KpnI
p299LYSA
KpnI digested
BamHI
KpnI
Brev.-ori
pHK4
BamHI digested
Blunt end
KpnI linker ligation
KpnI digested
Ligation
KpnI
lysA
Brev.-ori
KpnI
pLYSAB

FIG. 3

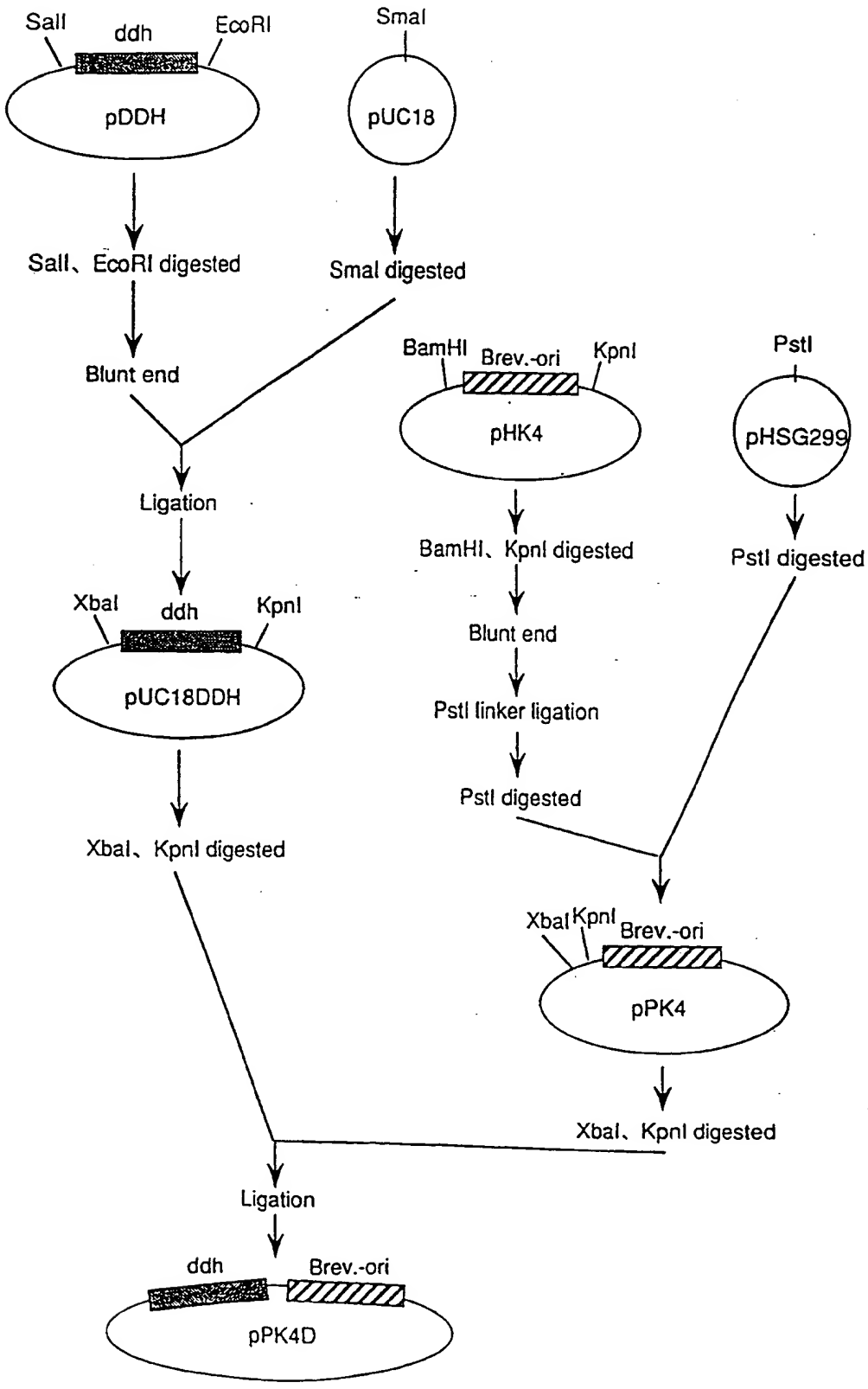

SalI
ddh
EcoRI
pDDH
SmaI
pUC18
SalI、EcoRI digested
SmaI digested
Blunt end
Ligation
BamHI
Brev.-ori
KpnI
pHK4
PstI
pHSG299
BamHI、KpnI digested
PstI digested
Blunt end
PstI linker ligation
PstI digested
XbaI
ddh
KpnI
pUC18DDH
XbaI、KpnI digested
XbaI
KpnI
Brev.-ori
pPK4
XbaI、KpnI digested
Ligation
ddh
Brev.-ori
pPK4D

FIG. 4

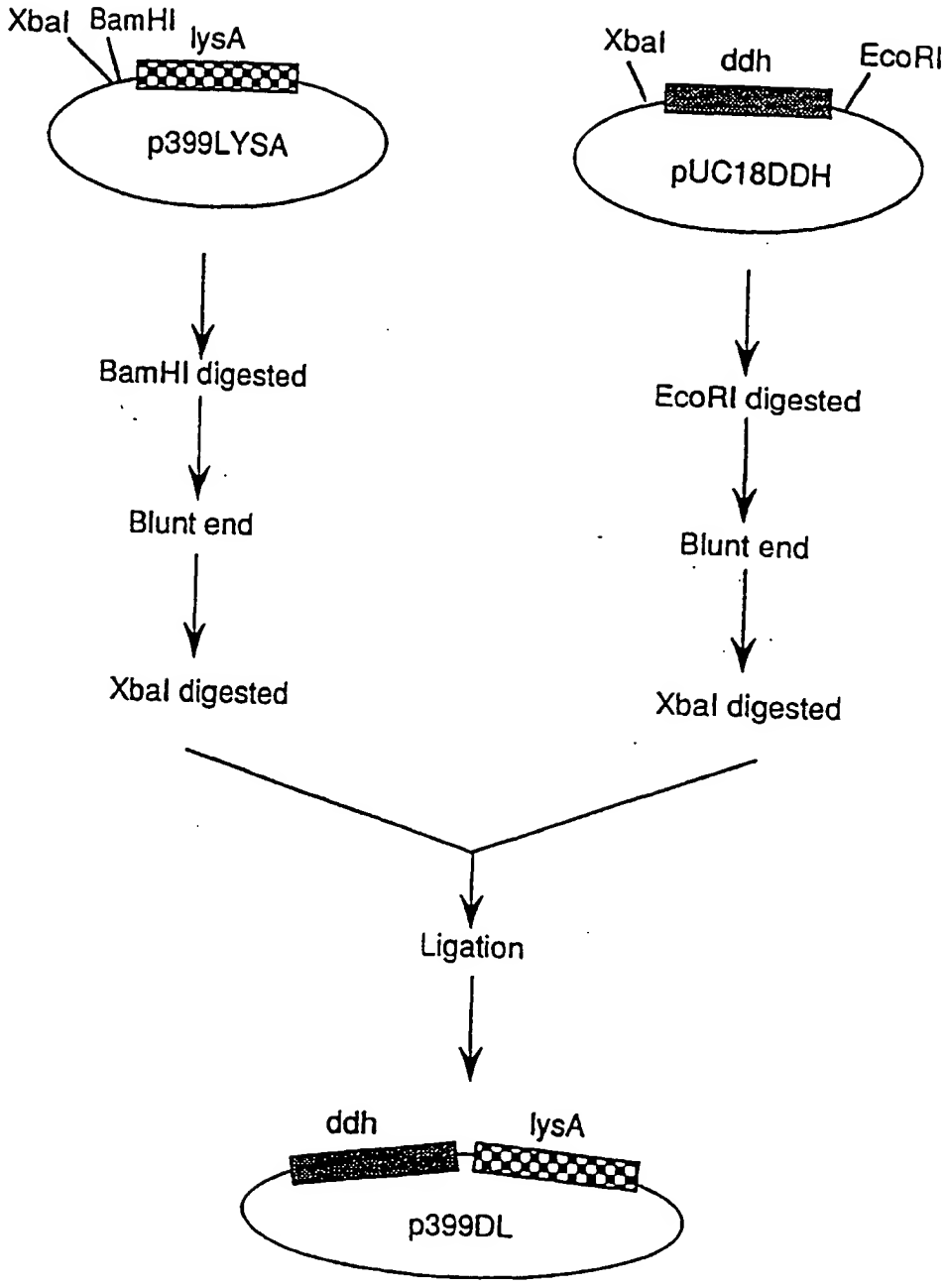

XbaI
BamHI
lysA
p399LYSA
BamHI digested
Blunt end
XbaI digested
XbaI
ddh
EcoRI
pUC18DDH
EcoRI digested
Blunt end
XbaI digested
Ligation
ddh
lysA
p399DL

FIG. 5

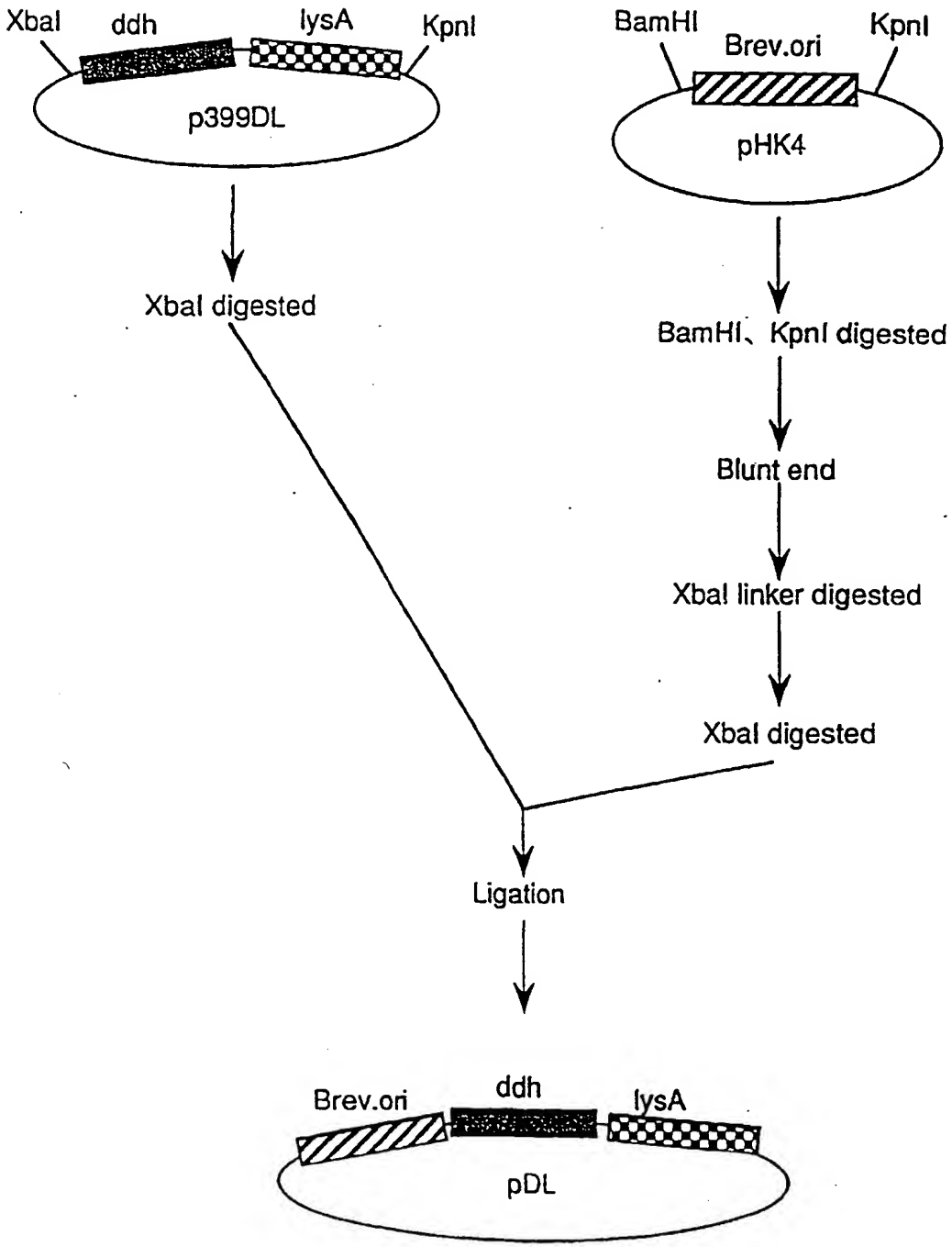

XbaI
ddh
lysA
KpnI
p399DL
XbaI digested
BamHI
Brev.ori
KpnI
pHK4
BamHI、KpnI digested
Blunt end
XbaI linker digested
XbaI digested
Ligation
Brev.ori
ddh
lysA
pDL

FIG. 6

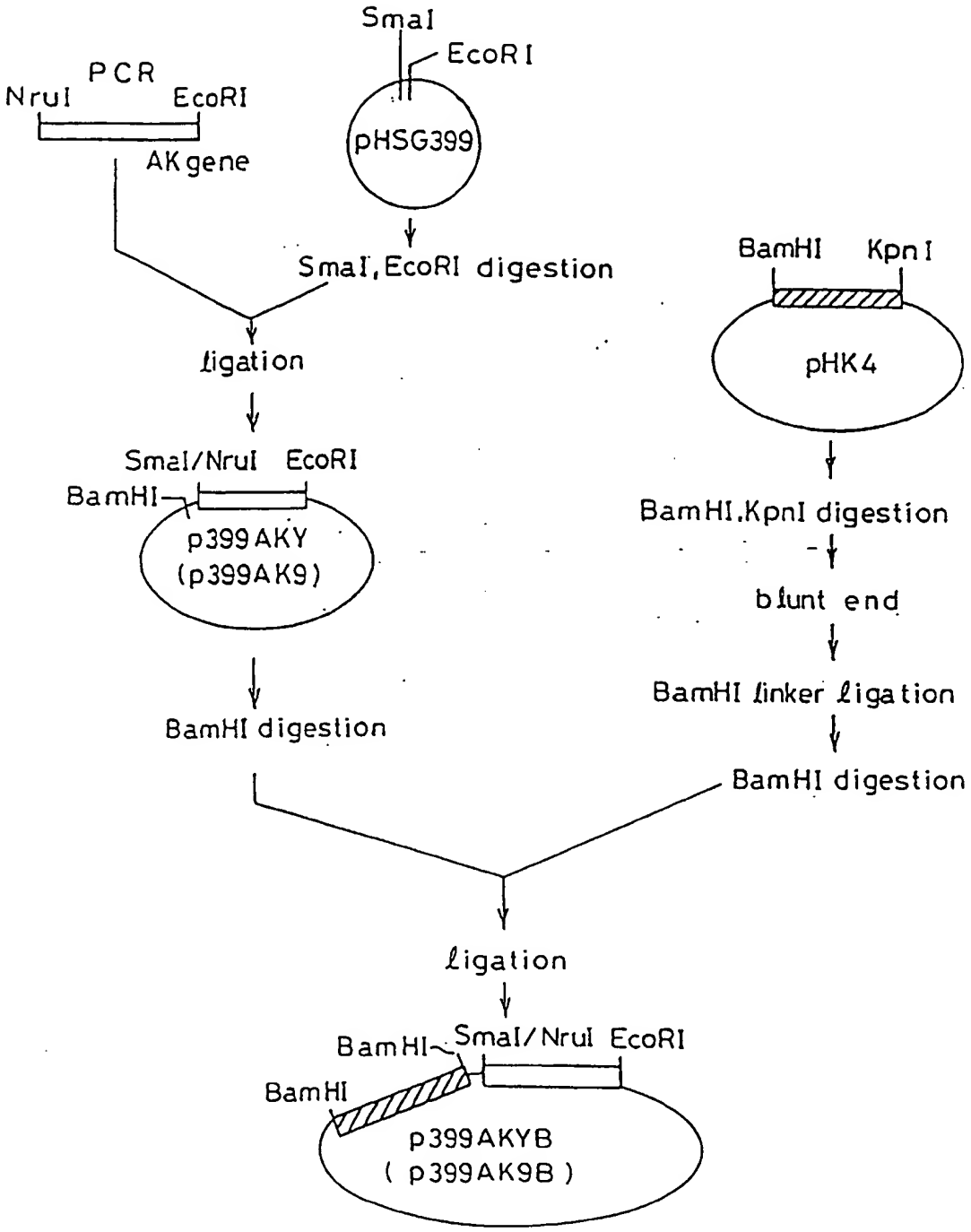

SmaI
EcoRI
PCR
NruI
EcoRI
AK gene
pHSG399
SmaI, EcoRI digestion
BamHI
KpnI
pHK4
ligation
SmaI/NruI
EcoRI
BamHI
p399AKY
(p399AK9)
BamHI, KpnI digestion
blunt end
BamHI linker ligation
BamHI digestion
BamHI digestion
ligation
BamHI
SmaI/NruI
EcoRI
BamHI
p399AKYB
(p399AK9B)

FIG. 7

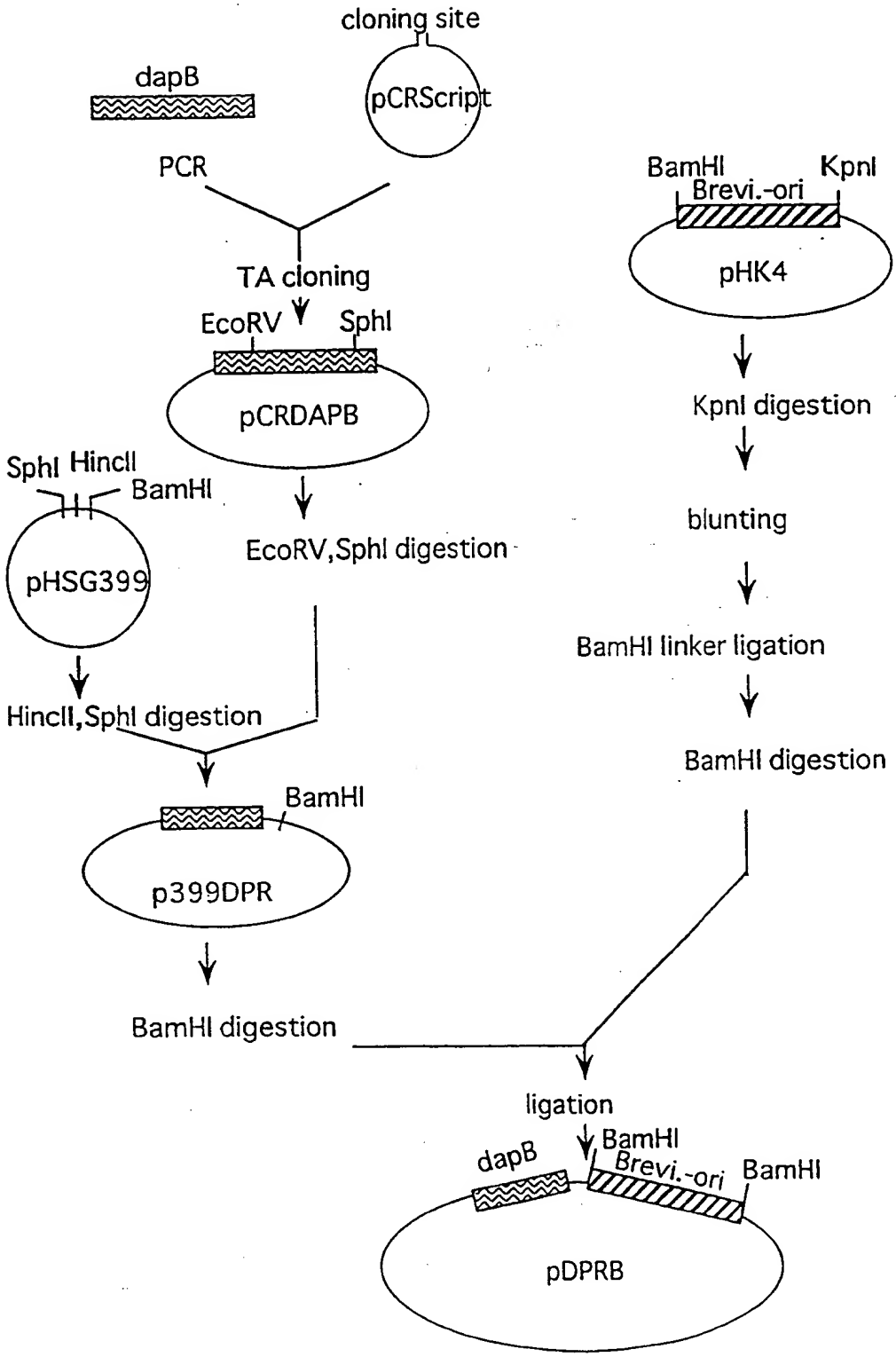

cloning site
dapB
pCRScript
PCR
TA cloning
EcoRV
SphI
pCRDAPB
SphI HincII
BamHI
pHSG399
EcoRV,SphI digestion
HincII,SphI digestion
BamHI
p399DPR
BamHI digestion
BamHI
Brevi.-ori
KpnI
pHK4
KpnI digestion
blunting
BamHI linker ligation
BamHI digestion
ligation
BamHI
dapB
Brevi.-ori
BamHI
pDPRB

FIG. 8

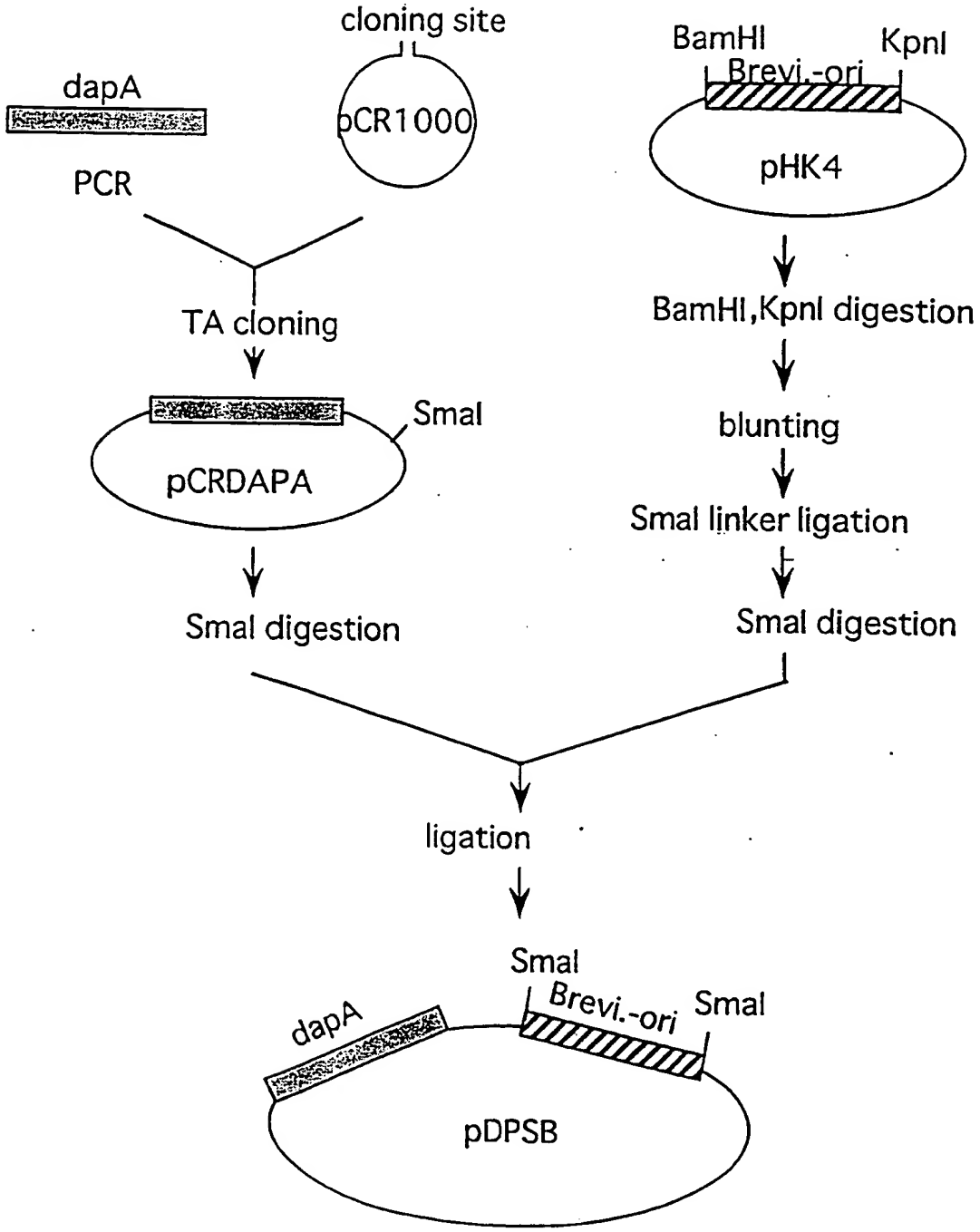

cloning site
dapA
pCR1000
PCR
TA cloning
SmaI
pCRDAPA
SmaI digestion
BamHI
KpnI
Brevi.-ori
pHK4
BamHI,KpnI digestion
blunting
SmaI linker ligation
SmaI digestion
ligation
SmaI
dapA
Brevi.-ori
SmaI
pDPSB

FIG. 9

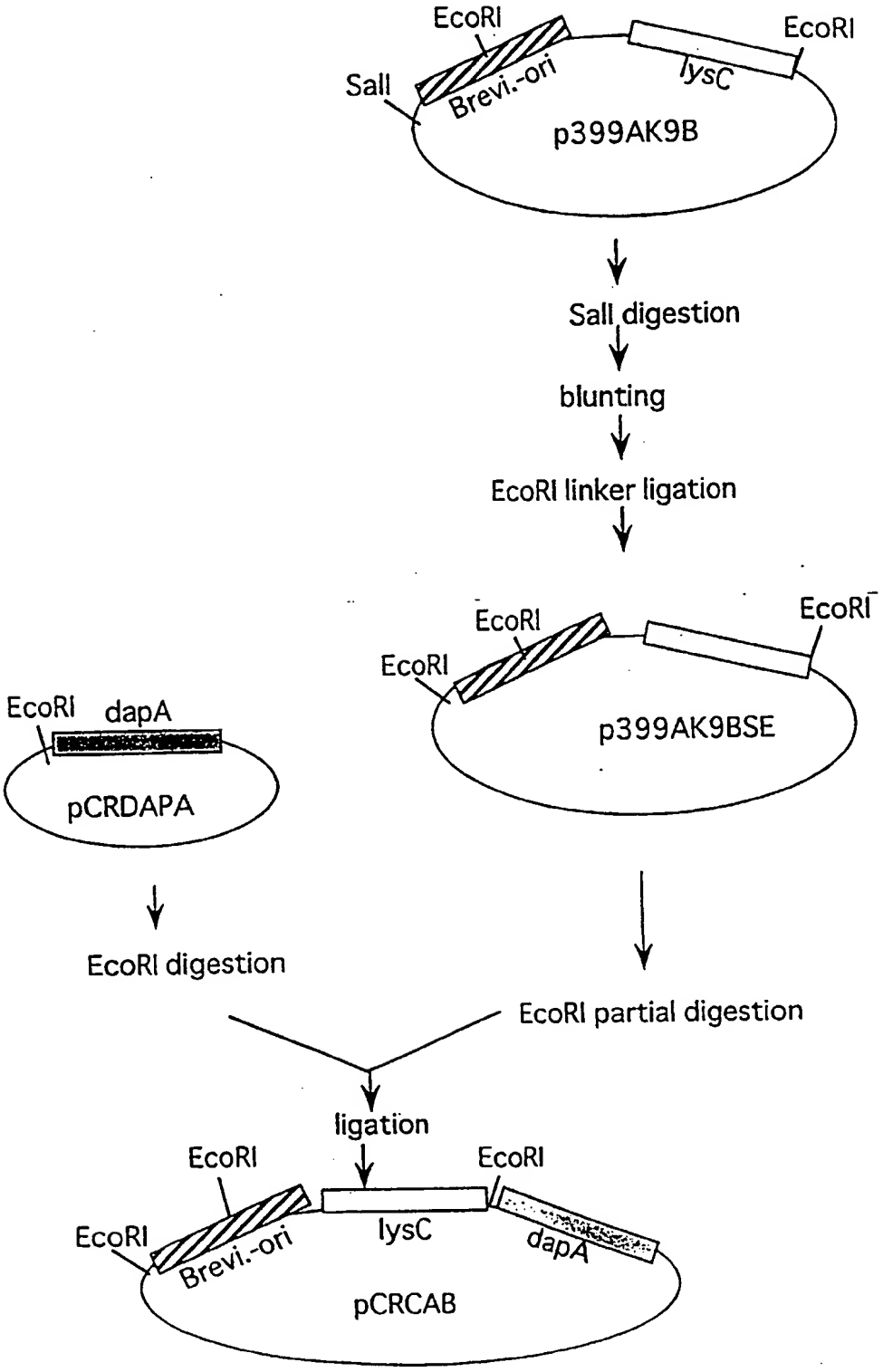

EcoRI
EcoRI
SalI
Brevi.-ori
lysC
p399AK9B
SalI digestion
blunting
EcoRI linker ligation
EcoRI
EcoRI
EcoRI
p399AK9BSE
EcoRI
dapA
pCRDAPA
EcoRI digestion
EcoRI partial digestion
ligation
EcoRI
EcoRI
EcoRI
Brevi.-ori
lysC
dapA
pCRCAB

FIG. 10

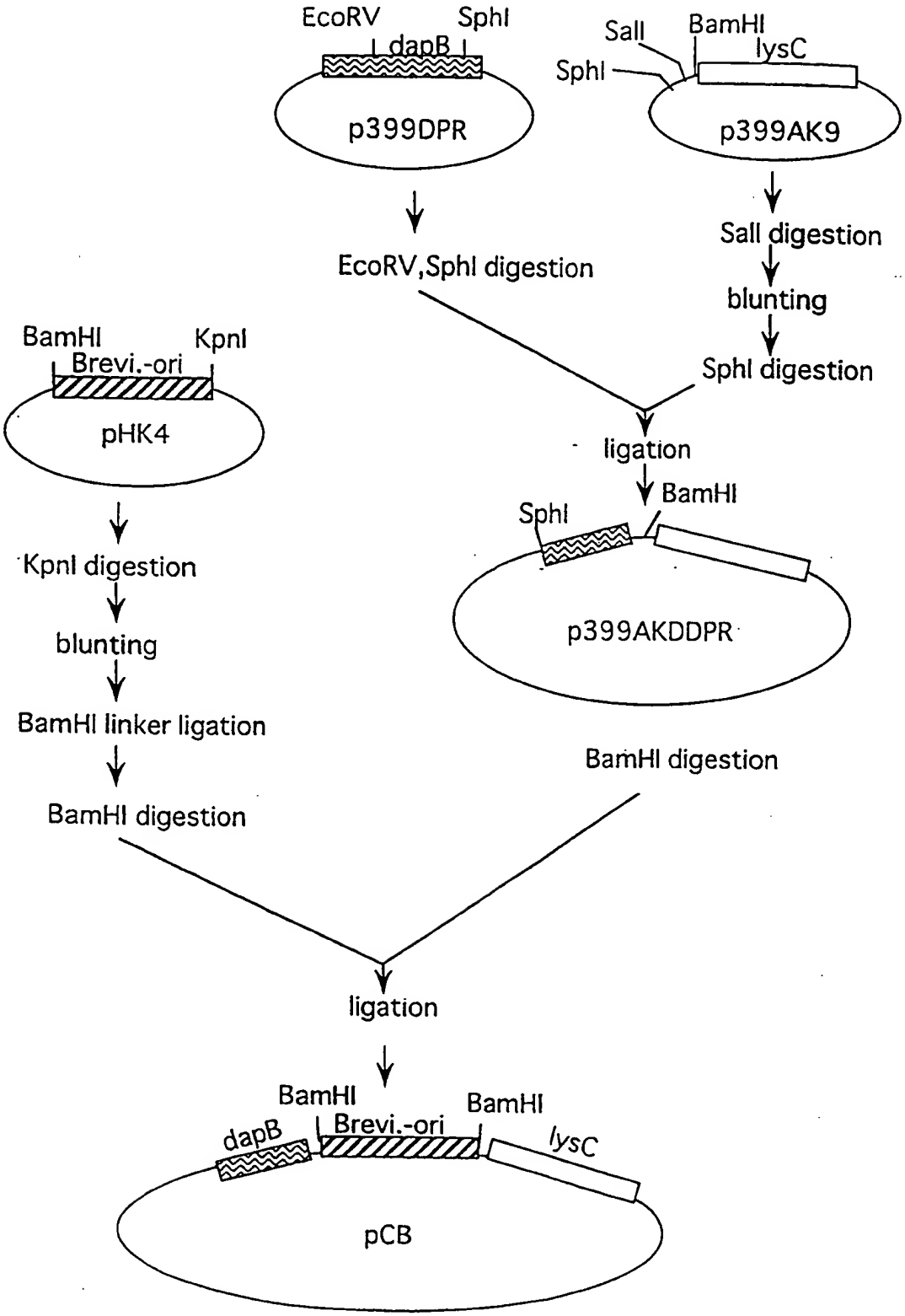

EcoRV
SphI
dapB
p399DPR
SalI
BamHI
lysC
SphI
p399AK9
SalI digestion
blunting
SphI digestion
EcoRV,SphI digestion
ligation
BamHI
SphI
p399AKDDPR
BamHI
Brevi.-ori
KpnI
pHK4
KpnI digestion
blunting
BamHI linker ligation
BamHI digestion
BamHI digestion
ligation
BamHI
BamHI
dapB
Brevi.-ori
lysC
pCB

FIG. 11

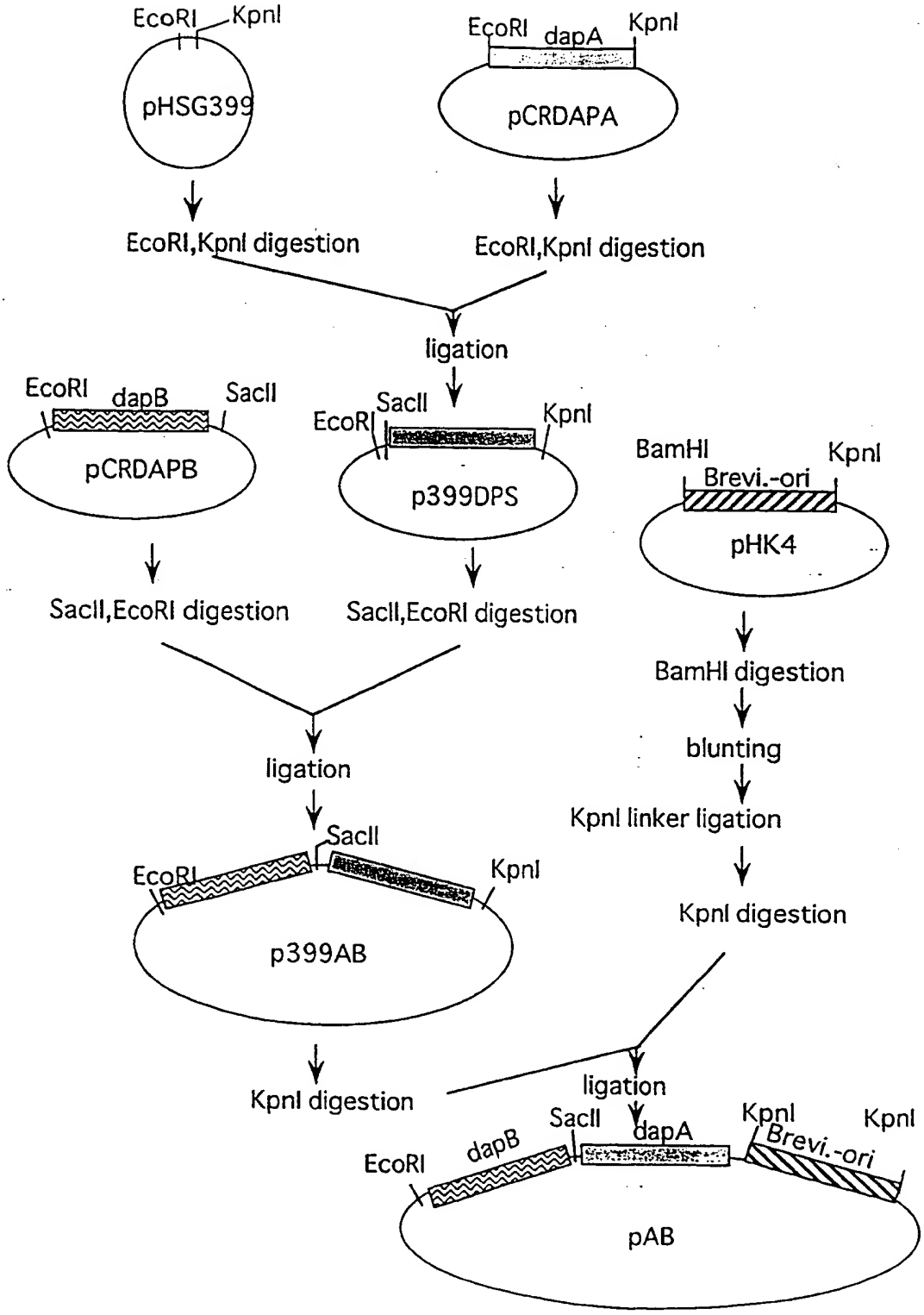

EcoRI
KpnI
pHSG399
EcoRI
dapA
KpnI
pCRDAPA
EcoRI,KpnI digestion
EcoRI,KpnI digestion
ligation
EcoRI
dapB
SacII
pCRDAPB
SacII
EcoRI
KpnI
p399DPS
BamHI
KpnI
Brevi.-ori
pHK4
SacII,EcoRI digestion
SacII,EcoRI digestion
BamHI digestion
blunting
KpnI linker ligation
ligation
SacII
EcoRI
KpnI
p399AB
KpnI digestion
KpnI digestion
ligation
SacII
KpnI
KpnI
EcoRI
dapB
dapA
Brevi.-ori
pAB

FIG. 12

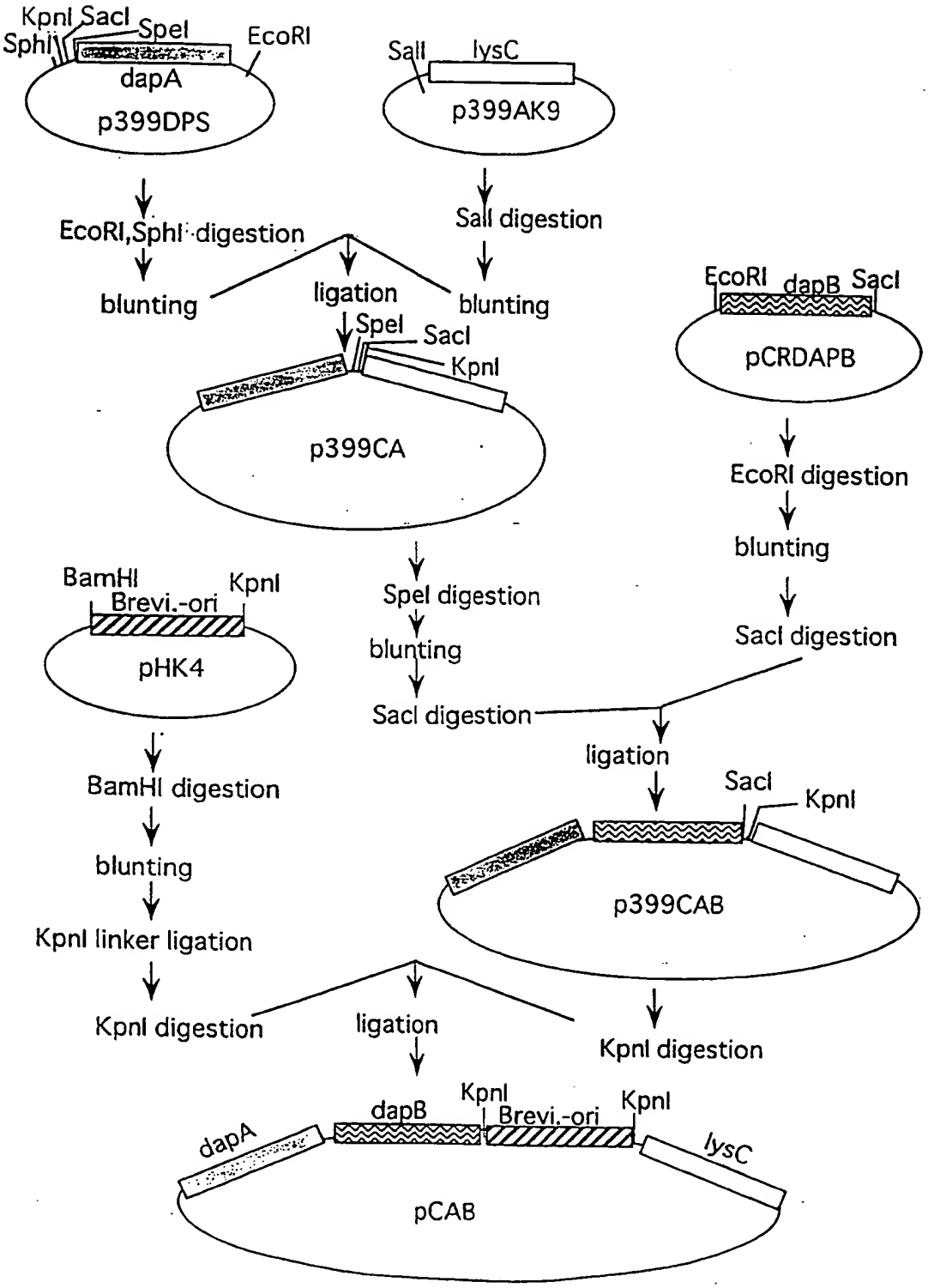

KpnI SacI
SphI
SpeI
EcoRI
dapA
p399DPS
SalI
lysC
p399AK9
EcoRI,SphI digestion
SalI digestion
blunting
ligation
blunting
SpeI
SacI
KpnI
p399CA
EcoRI
dapB
SacI
pCRDAPB
EcoRI digestion
blunting
SacI digestion
BamHI
KpnI
Brevi.-ori
pHK4
SpeI digestion
blunting
SacI digestion
ligation
SacI
KpnI
p399CAB
BamHI digestion
blunting
KpnI linker ligation
KpnI digestion
ligation
KpnI digestion
KpnI
KpnI
dapB
Brevi.-ori
dapA
lysC
pCAB

FIG. 13

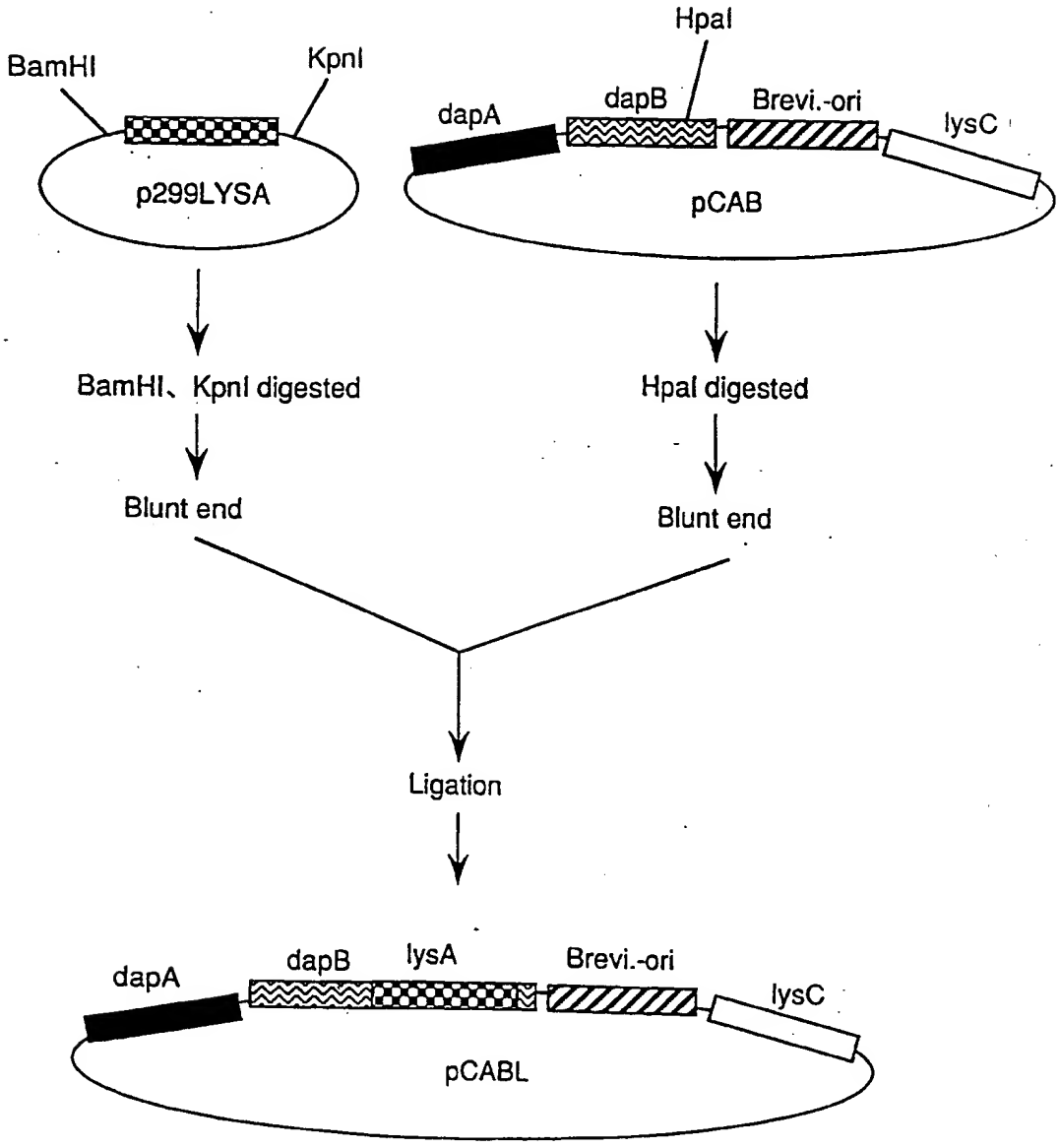
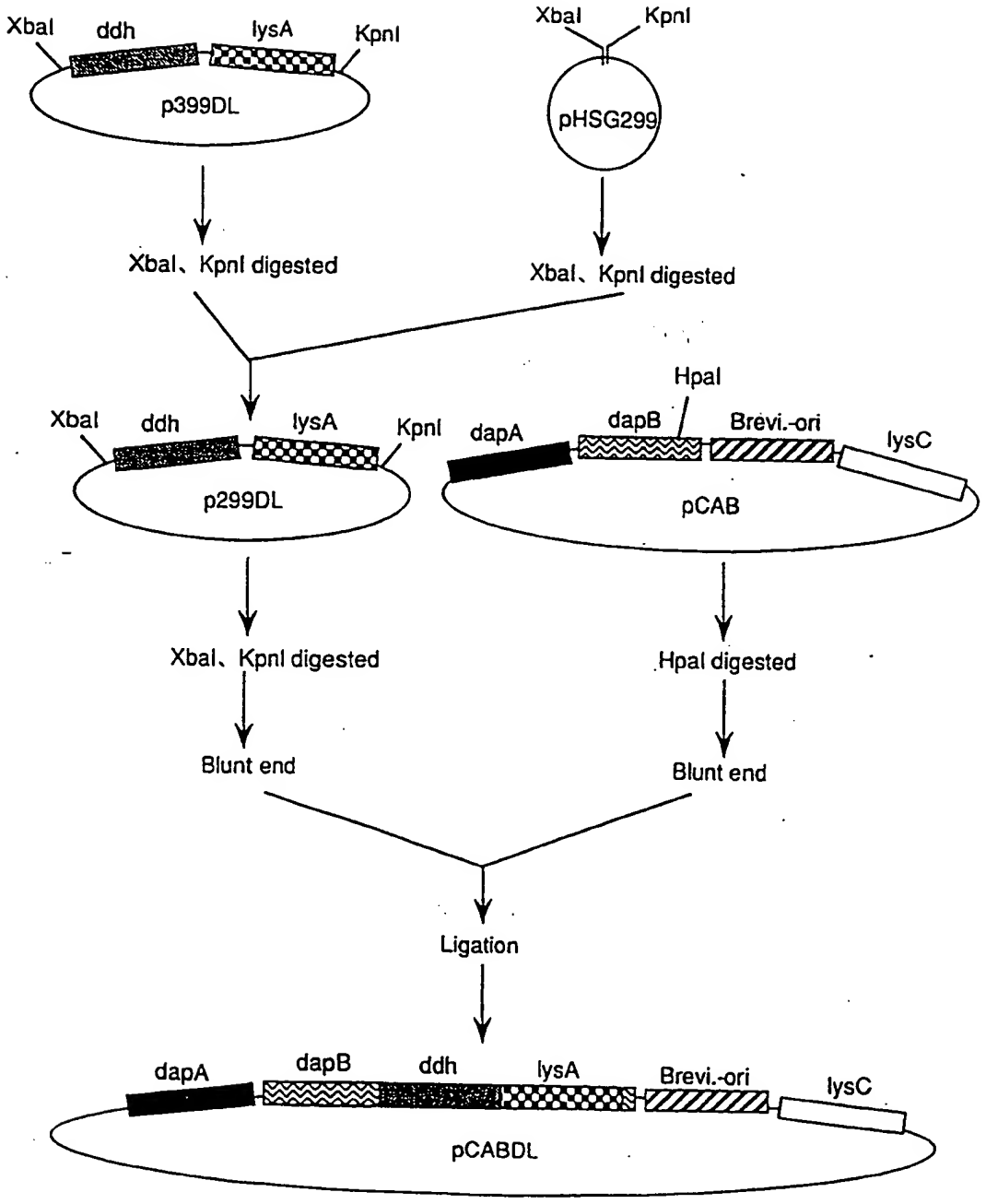

BamHI
KpnI
p299LYSA
HpaI
dapA
dapB
Brevi.-ori
lysC
pCAB
BamHI、KpnI digested
HpaI digested
Blunt end
Blunt end
Ligation
dapA
dapB
lysA
Brevi.-ori
lysC
pCABL

FIG. 14


XbaI
ddh
lysA
KpnI
p399DL
XbaI
KpnI
pHSG299
XbaI、KpnI digested
XbaI、KpnI digested
HpaI
XbaI
ddh
lysA
KpnI
dapA
dapB
Brevi.-ori
lysC
p299DL
pCAB
XbaI、KpnI digested
HpaI digested
Blunt end
Blunt end
Ligation
dapA
dapB
ddh
lysA
Brevi.-ori
lysC
pCABDL

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